

The Protective Effects of *Helicobacter pylori* Infection on Allergic Asthma

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Keywords

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Abstract

As an ancient Gram-negative bacterium, *Helicobacter pylori* has settled in human stomach. Eradicating *H. pylori* increases the morbidities of asthma and other allergic diseases. Therefore, *H. pylori* might play a protective role against asthma. The “disappearing microbiota” hypothesis suggests that the absence of certain types of the ancestral microbiota could change the development of immunology, metabolism, and cognitive ability in our early life, contributing to the development of some diseases. And the Hygiene Hypothesis links early environmental and microbial exposure to the prevalence of atopic allergies and asthma. Exposure to the environment and microbes can influence the growing immune system and protect subsequent immune-mediated diseases. *H. pylori* can inhibit allergic asthma by regulating the ratio of helper T cells 1/2 (Th1/Th2), Th17/regulatory T cells (Tregs),

etc. *H. pylori* can also target dendritic cells to promote immune tolerance and enhance the protective effect on allergic asthma, and this effect relies on highly suppressed Tregs. The remote regulation of lung immune function by *H. pylori* is consistent with the gut-lung axis theory. Perhaps, *H. pylori* also protects against asthma by altering levels of stomach hormones, affecting the autonomic nervous system and lowering the expression of heat shock protein 70. Therapeutic products from *H. pylori* may be used to prevent and treat asthma. This paper reviews the possible protective influence of *H. pylori* on allergic asthma and the possible application of *H. pylori* in treating asthma.

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Introduction

Since Gram-negative *Helicobacter pylori* was discovered, the relationship between *H. pylori* infection and asthma has gradually attracted people’s eyes. Kalach et al. [1] analyzed the infection of *H. pylori* in adults and chil-

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dren from the perspective of host response, epidemiology, related diseases, clinical features, therapies, and diagnosis. The incidence of *H. pylori* infection is decreasing in both adults and children in developed countries and several developing areas, which is opposite from the increased incidence of asthma in children and other allergic disorders. Ness-Jensen et al. [2] found that *H. pylori* infection has been related to a 40% decrease of asthma in those who are below the age of 18 years with abdominal obesity. A recent study found that 16.4 percent of children who were negative for *H. pylori* at the age of 2 and 10 had asthma at the age of 16, but if they were positive for *H. pylori* at the age of 12, they did not have asthma at the age of 16. It is suggested that the early exposure to *H. pylori* can prevent asthma [3]. Fouda et al. [4] used ELISA for determination of *H. pylori* IgG in the serum of asthma and healthy children. The results showed that the titer of IgG was negatively correlated with the degree of asthma. Serum *H. pylori* could prevent asthma in children and was inversely correlated with the severity of asthma. Accumulating evidence suggested that the infection of *H. pylori*, particularly CagA-positive *H. pylori*, was negatively correlated with the development of asthma. A study of Greek children has also proved this viewpoint [5]. Recent and earlier cross-sectional studies also suggested that *H. pylori* infection had protective effects on asthma [6–9]. However, not all studies supported this result, which still requires future research [10–16]. Although the effects of *H. pylori* in the pathophysiological mechanisms of asthma still remain controversial, the researchers detected *H. pylori* exotoxin VacA in human lung biopsies and directly stimulated pulmonary airway epithelial cells to secrete inflammatory cytokines in vitro [17]. Moreover, *H. pylori* has been discovered in the lung tissues of patients with COPD [18]. More studies confirmed that existence of *H. pylori* provided protective effects against asthma, and eradication of *H. pylori* may have a negative impact [19–21].

The immune system includes adaptive immunity (acquired immunity) and innate immunity (natural immunity) [22–24]. Adaptive immunity mainly recognizes “non-self” antigen and produces immune tolerance, and innate immunity is the first line of defense against pathogenic microorganism invasion, which can effectively distinguish self from pathogenic microorganism [25, 26]. The immune pathogenesis of allergic asthma is quite complex. The studies have focused on Toll-like receptors (TLRs), dendritic cells (DCs), helper T cells 1/2 (Th1/Th2), Th17, regulatory T cells (Tregs), etc. The formation of a complex interaction network between cells and re-

ceptors also provides a broad view for immunological research of asthma [27–29]. The followings are the possible protective mechanisms of *H. pylori* against allergic asthma reported in recent years and the possible associated treatment of asthma.

Possible Protective Mechanisms of *H. pylori* against Allergic Asthma

The Hygiene Hypothesis

The hygiene hypothesis [30, 31] links early environmental and microbial exposure to the prevalence of atopic allergies and asthma. The exposure to environment and microbes can help to form the growing immunity system and protect subsequent immune-mediated diseases [32]. “Unhygienic exposure” to microorganisms in an early age can prevent the development of allergic diseases in later years [33]. *H. pylori* infection usually occurs in children, and the way to be infected is related to unhygienic family environment or habits, and this association appears in mouse asthma models [34]. Early studies [35] have shown that the hygiene hypothesis may be related to Th1/Th2 imbalance. Synthetic adjuvants or microbial components can directly influence the cells in the innate immune system, including NK and DCs cells, and also stimulate the secretion of interferon- γ (IFN- γ), IL-12, and IFN- α , leading to the phenotypic transformation of allergen-specific Th2 to Th1 cells [7, 36]. The specific mechanisms still need to be further studied.

Adjusting Th1/Th2 Balance

It has been proved that Th1/Th2 ratio imbalance is one of the essential immunological mechanisms of asthma. According to the responses to foreign antigens, T cells can be divided into 2 types of effector cells, Th1 and Th2, which have totally different functions [37, 38]. Th1 mainly secretes IL-12, IFN- γ , and transforming growth factor β (TNF- β), activates macrophages and causes cytotoxicity, and mediates cellular immunity. Th2 mainly secretes IL-4, IL-5, and IL-13, activates B cells to produce immunoglobulin, and mediates humoral immunity. Th1 and Th2 are restrictive to each other and reach a balance. Asthma is a disease characterized by the count of Th2 and the effects it exerts [39–41].

H. pylori neutrophil-activating protein (HP-NAP) is one of the main virulence factors of *H. pylori*, which is also applied as a possible biomarker in the diagnosis of *H. pylori*-related diseases [42]. Studies [43–45] have shown that HP-NAP plays a protective role in asthma, which

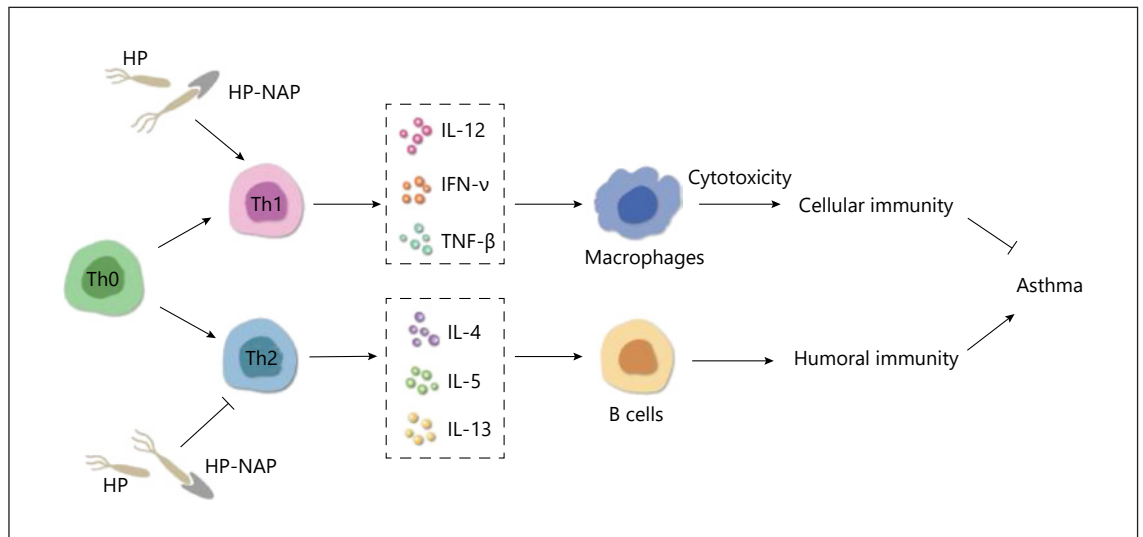


Fig. 1. *H. pylori* promotes Th1 activation and inhibits Th2 response through HP-NAP in allergic asthma. HP, *H. pylori*; HP-NAP, *H. pylori* neutrophil-activating protein; Th0/Th1/Th2, helper T cells 0/1/2.

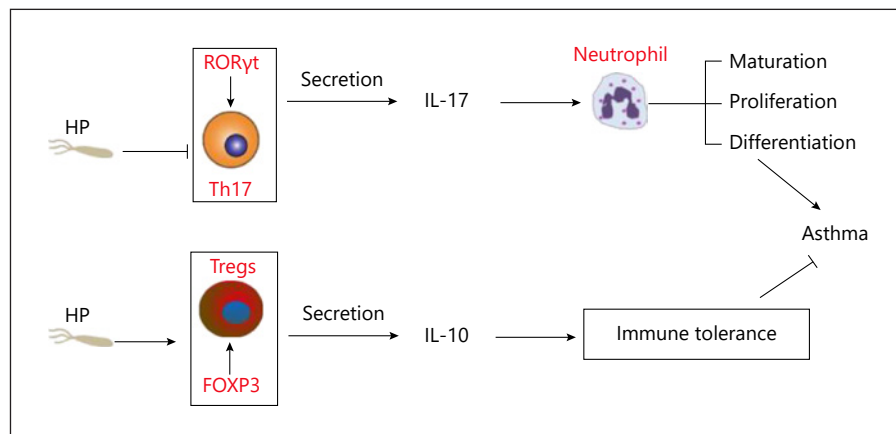
could stimulate Th1 activation and attenuate Th2 response in allergy-related asthma both in vitro and in vivo (Fig. 1). In the research by Karakullukcu et al. [46], 18 cases (20.4%) of *H. pylori* DNA were discovered in 88 healthy stool samples but none in 92 asthmatic children (3–8 years). Multivariate Logistic regression analysis suggested that HP-NAP had a protective effect on asthma in male children. In order to verify HP-NAP as a regulatory factor against the Th2 inflammatory effect, Zhou et al. [47] exposed the mice to purified recombinant *H. pylori* NAP (rNAP) through intraperitoneal injection or inhalation. The increase and infiltration of the eosinophils were remarkably suppressed in the lungs of the asthma mice model induced by ovalbumin (OVA). Moreover, the count of eosinophils was decreased in the lavage fluid from bronchoalveolar (BALF) in the mice treated with rNAP. Additionally, the levels of IL-13 and IL-4 declined ($p < 0.01$), the levels of IFN- γ and IL-10 elevated ($p < 0.01$), and the serum level of IgE declined ($p < 0.01$) in experimental groups in comparison with the control group. It is suggested that mucosal and systemic pretreatment of rNAP might attenuate asthma in the mice induced by OVA. Furthermore, rNAP could be used as a new method in preventing or treating allergic disorders. In phase I/II clinical trials, it has been reported that the effect of soluble IL-4 receptor (sIL-4R) was satisfactory in treating asthma. To discover more effective antiasthma therapies, Liu et al. [48] attempted to test whether the HP-NAP could increase the therapeutic effect of sIL-4R on

asthma. In their research, pcDNA3.1-sIL-4R-NAP plasmid (PSN) was constructed, which could encode a fusion protein of murine HP-NAP and sIL-4R. PSN could remarkably reduce inflammation in the airway, suppress the secretion of OVA-specific IgE in serum, and reestablish the balance of Th1 and Th2. Moreover, PSN has been more effective in the treatment for asthma in comparison with the plasmid only generating sIL-4R.

Adjusting Th17/Tregs Balance

With the development of scientific research, Th1/Th2 imbalance cannot fully explain the mechanisms of asthma [49]. Although allergic asthma is often associated with abnormal TH2 cellular responses, a group of patients with severe disease showed a mixture of TH2 and TH17 cellular responses in the airways [50]. It was found that Th17 and Tregs cells were also significantly related to the pathogenesis of asthma [51–53]. Synergy of multiple pathways, such as Th2, Th17, and even eosinophil/neutrophil infiltration, has been found in some asthma models [54–56]. The view that eosinophilic asthma is an exclusive TH2 disorder and neutrophil asthma is an exclusive TH17 disorder may be oversimplified [57]. It has been found that the TH2 and TH17 inflammatory pathways regulate each other in asthma [58]. Th1/Th2 and Th17/Tregs and their various cytokines form an extremely complex interactive network [59, 60]. Th17 cells are defined as “proinflammatory” immune cells, which mainly secrete IL-17, mediate inflammatory responses, and pro-

Fig. 2. *H. pylori* has a protective effect on asthma by inducing highly immunosuppressive Tregs and inhibiting Th17. HP, *H. pylori*; Tregs, regulatory T cells; ROR γ t, retinoid-related orphan receptor (ROR) gamma t; Foxp3, forkhead transcription factor p3.



mote neutrophil maturation, proliferation, and chemotaxis. Retinoid-related orphan receptor (ROR) gamma t (ROR γ t) is a key transcriptional factor in the differentiation of Th17 cells. The deletion of ROR γ t leads to the failure of Th17 differentiation [61, 62]. Tregs secrete IL-10 and other inhibitory cytokines. Tregs play an important role in maintaining immune balance. Tregs specifically express forkhead transcription factor p3 (Foxp3), suppress immune response, and mediate immune tolerance [63–65].

The protective mechanisms of *H. pylori* in inflammatory diseases such as asthma may be associated with induction of Tregs, which could highly suppress the immune activity [66–68]. Published data strongly suggested that *H. pylori* caused an increase in the response levels of Tregs, Th1, and Th17 in mouse models and human, which could prevent asthma. According to the ontogeny of the immune system, *H. pylori* tends to infect in one's childhood and continues to stimulate immunity reaction throughout the life, including Th1, Th17, and Tregs responses. Additionally, children infected by *H. pylori* tend to have a stronger Tregs response than adults [69–72]. Once human is infected by *H. pylori*, Tregs are found to be highly active in the gastric mucosa. The secretion of IL-10 by Tregs in peripheral blood was significantly higher than that in *H. pylori*-specific Th1 cells. When there was a strong Tregs reaction, the concentration of total IgE and allergen-specific IgE was low. Suppressing IL-10 could significantly restore the IgE reaction in animal models. Therefore, systematic IL-10 and Tregs may play a role in preventing allergies mediated by *H. pylori* [72].

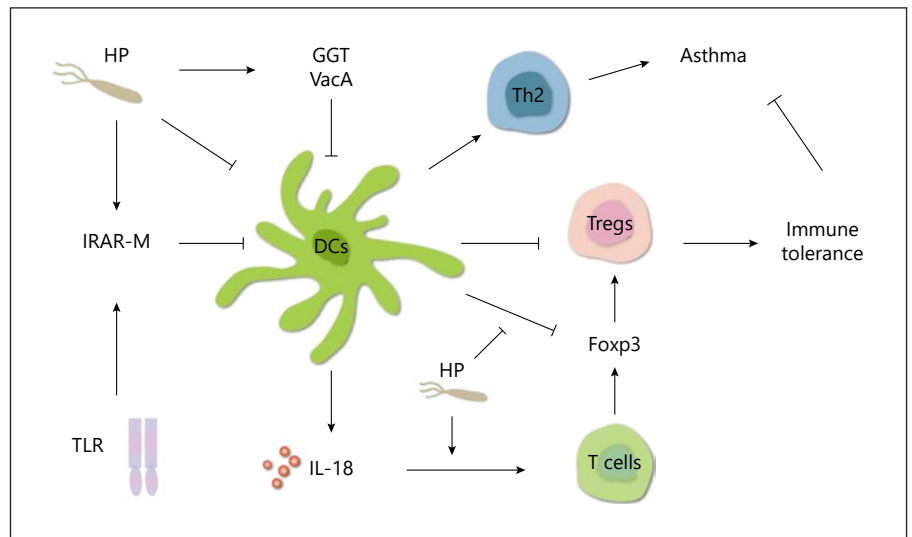
Kyburz et al. [73] established C57BL/6 mice experimental models of house dust mite- or ovalbumin-induced airway inflammation and influenza A virus or *Citrobacter*

rodentium infection. It was found that the exposure to *H. pylori* extract or its immunomodulator vacuolating cytotoxin in the perinatal stage could exert robust protective functions against allergic inflammation in the airway not only in the offspring of the first generation but also the second generation, which did not increase the susceptibility to bacterial or viral infection. The immune responses correlated with prevention of allergy include inhibiting the activities of effectors or T cells, expanding the subsets of regulatory T cells expressing ROR γ t, and FOXP3 demethylation. The diversity and composition of the microbiota in the gastrointestinal system are notably influenced by the perinatal exposure of *H. pylori*. In conclusion, *H. pylori* exposure not only works on the carriers but also on the next generations. Maternal nutrient, the exposure to microorganisms, tobacco, and other environmental factors influence the formation of the immune system in a fetus via an epigenetic way (Fig. 2).

Inhibition of DCs

DCs can devour and kill invasive microorganisms and present microbial antigens to T cells, thus participating in the innate immunity [25, 74]. DCs help the initial T cells polarize into Th2 effector cells or differentiate into Tregs [75, 76]. It has been suggested that *H. pylori* targets DCs to promote immune tolerance and enhance the protective effect against allergic asthma, and this effect relies on highly suppressed Tregs [77, 78]. Oertli et al. [79, 80] found the level of Foxp3, the main regulatory factor expressed by Tregs, was increased in immature T cells when *H. pylori* was exposed to DCs in cells and animal experiments. Depleting the DCs in *H. pylori*-infected mice in the neonatal stage resulted in the improvement of infection control and the destruction of specific tolerance to *H*

Fig. 3. *H. pylori* targets DCs and relies on highly suppressed Tregs to promote immune tolerance and enhance the protective effect on allergic asthma. HP, *H. pylori*; Tregs, regulatory T cells; DCs, dendritic cells; TLR, Toll-like receptors; GGT, γ -glutamyl transpeptidase; VacA, vacuolating cytotoxin A.



pylori. At the same time, it also aggravated the immunopathological reactions driven by T cells. IL-18 secreted by DCs could directly act on T cells and promote transformation into Tregs. It promotes specific immune tolerance and asthma protection in mice. Vacuolating cytotoxin A (VacA) and γ -glutamyl transpeptidase (GGT), 2 virulence factors in *H. pylori*, independently interfere with the maturation of DCs, thus promoting the tolerance of DCs. Shiu et al. [81] found that the infection of *H. pylori* can upregulate the expression of anti-inflammation factors including IL-1 receptor-associated kinase M (IRAK-M). The expression of IRAK-M activated by TLRs in DCs directly inhibited the inherent function of DCs, such as the upregulation of cytokines and costimulatory molecules, rather than affecting the response of Th17 and Tregs (Fig. 3).

Activation of TLRs

TLRs are a group of widely studied pattern recognition receptors, which are associated with the incidence and progression of asthma. At present, it is considered that TLRs are mainly expressed in the membrane or organelle capsule of antigen presenting cells (such as macrophages and DCs). TLRs participate in the early host defense and play an important role in the innate immune response. On the other hand, TLRs participate in the inflammatory response via secreting cytokines, chemokines, adhesion molecules, etc., and finally activate the acquired immune system. Different TLRs have different functions [82–84]. TLR2 is an important pattern recognition receptor in Tregs [85, 86] and TLR9 mainly plays a negative role in

the regulation of allergic inflammation [87–89]. TLR4 not only relies on conserved sequences encoded by embryological genes to identify pathogenic microorganisms and activate innate immunity but also regulates adaptive immunity [90, 91]. In conclusion, TLRs are closely related to asthma [92].

In recent days, hypotheses were proposed that *H. pylori* could facilitate the activation of inflammasomes in mouse and human immune cells. The possible mechanisms and virulence factors stimulating the inflammasome have been discovered in animal and cell models. IL-1 β could facilitate the responses of Th1 and Th17. IL-18 has been a hallmark in humans and mice infected by *H. pylori*, which plays an important role in *H. pylori* persistence, Tregs differentiation, and prevention of asthma. The secretion of IL-1 β induced by *H. pylori* is regulated by the activation of NLRP3 (Nod-like receptor family member), caspase-1, and TLR2. The axis of TLR2/NLRP3/caspase-1/IL-18 was essential in the regulation of *H. pylori*-specific immune response which could prevent inflammatory bowel disease and asthma induced by allergens in mouse models [93–95].

NOD1 is considered as a pattern recognition receptor in cells and specifically targets the Gram-negative peptidoglycan, which plays an important role in host defense against infections (e.g., *H. pylori* and *Shigella flexneri*) [96]. The variations in the NOD1 gene contribute to inflammatory bowel disease and asthma. NOD1 could be activated by a rather low concentration of M-Tri DAP, which is a specific muropeptide ligand. Moreover, NOD1 could induce minimal secretion of IL-10, TNF- α , and IL-1 β from

peripheral blood mononuclear cells in human and synergistically facilitated the responses induced by TLRs. Synergistic responses occurred across a variety of cytokine secretions (GM-CSF, IL-10, IL-4, IL-6, IL-1 β , IL-1 α , and TNF- α) and various ligands (to TLR5, 7/8, 2/6, 1/2, 4) [97].

TLR9 could partly contribute to the initiation of immunity responses induced by bacteria via binding to the unmethylated CpG-DNA rich in bacteria. A well-reported single nucleotide polymorphism (SNP) in the TLR9 promoter (TLR9-1237T/C) is related to multiple inflammatory diseases, such as atopy, allergic asthma, and inflammatory bowel disease. The sequence of the TLR9 promoter gene was analyzed, and the results demonstrated that carrying the variant “C” allele on position -1237 formed a possible site to bind NF- κ B, which could theoretically stimulate the transcriptional process of the gene [98]. It has also been shown that the type IV secretion system of *H. pylori* facilitated the synthesis of IL-8 through the p38 protein kinase (p38MAPK) pathway in the primary tracheobronchial epithelial cells collected from young rhesus monkeys. It was suggested that the innate immune response in airway epithelial cells in infants infected by *H. pylori* was enhanced, but the TLR4 pathway was not essential in this process [99].

Reduction of Gastroesophageal Reflux

Gastroesophageal reflux can induce or aggravate asthma. Several possible mechanisms have been raised. The first is stimulation theory. The airway is stimulated by the aspiration resulting from the reflux, which increases airway responsiveness. The second is reflex theory. Since esophagus and bronchus are derived from the same embryonic organ, the autonomic innervation is similar. Reflux not only stimulates esophagus but also activates the vagus nerve, thus inducing bronchospasm and aggravating asthma [100, 101]. It has been shown that *H. pylori* inflammation changed gastric hormonal status and influenced the autonomic nervous system. *H. pylori* can also reduce gastroesophageal reflux [102].

The Gut-Lung Axis Theory

In recent decades, the role of gut flora in the pathogenesis of asthma has been extensively studied [103–105]. The gut and lungs interact with each other through microbes and immune functions, achieve bidirectional regulation, and amplify immune signals. It is known as the gut-lung axis [106]. Gut and lung microbes have certain homology at early colonization. They all first pass through the oropharynx and then enter the digestive tract or respiratory tract through swallowing or breathing [107].

Microorganisms in the gastrointestinal tract can reach the lower respiratory tract through gastroesophageal reflux [108]. Due to the increase of intestinal and alveolar capillary permeability in some patients, the bacteria from the intestinal mucosa can be transferred to the lungs through the blood circulation [109]. Changes in the pulmonary flora can also cause changes in the intestinal flora through the blood flow [110]. However, there is little evidence about direct shift of microorganisms between 2 sites [107].

Disorders of the gut can be observed in lung diseases [105, 111–114]. Influenza virus can change the composition of intestinal flora and cause intestinal immune damage through Th17 cell mediation [115]. Locally induced pulmonary anaphylaxis may also affect the composition of intestinal flora [116]. Studies have shown that the severity of intestinal symptoms is highly consistent with the severity of pulmonary symptoms [117, 118].

Intestinal microbes promote development of the body's immune system early in life and affect the whole body and lungs through the blood and lymphatic system [104]. Both the gut and lungs have a strong mucosal defense system. For instance, intestinal and respiratory mucosal goblet cells can secrete IgA. The intestinal microflora can regulate pulmonary immune responses through bacterial lipopolysaccharide, short-chain fatty acids (SCFAs), and immune cells (e.g., Tregs and DCs), which can affect colonization of the lung microbiome [106, 119, 120]. The imbalance of intestinal flora is related to a variety of lung diseases such as asthma. The adjustment of intestinal flora can alleviate the symptoms and reduce the incidence of asthma. Probiotics supplementation has a certain preventive and therapeutic effect on asthma in mice. High-fiber diet can change the intestinal flora of mice and increase the content of SCFAs, thus inhibiting the activity of Th2 cells [121]. *H. pylori* can cause chronic immunopathologic changes in the stomach and dysbacteriosis and promote regulation of the immune function of the lung. *H. pylori* protects asthma by DCs, Tregs, etc., which is consistent with the gut-lung axis theory [106, 110, 122] (Fig. 4).

Reducing the Expression of Heat Shock Protein 70

Heat shock protein 70 (HSP70), an ATP-dependent chaperone protein, is a known inhibitor of caspase activation, showing antiapoptotic activity in a variety of cells [123, 124]. It has been found that HSP70 might play a role in promoting asthma inflammation. HSP70 deficiency leads to significant reduction in airway inflammation, goblet cell proliferation, and Th2 cytokine production, including IL-4, IL-5, and IL-13, and targeting HSP70 can

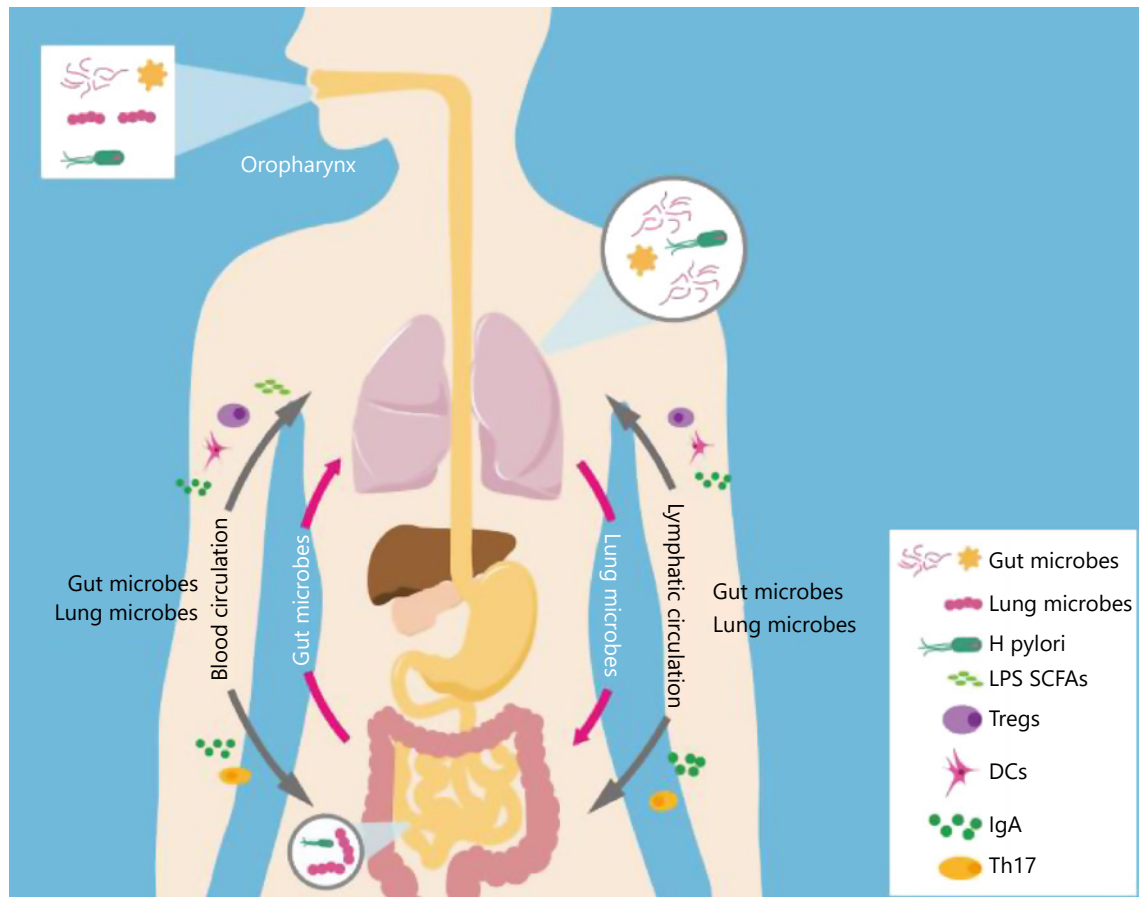


Fig. 4. The gut-lung axis theory: Gut and lung interactions through microbiota and immune function. LPS, bacterial lipopolysaccharide; SCFAs, short-chain fatty acids; Tregs, regulatory T cells; DCs, dendritic cells; Th17, helper T cells 17.

alleviate the potential utility of allergen-induced Th2 cytokines, goblet cell proliferation, and airway inflammation [125–127]. HSP70/CD80 DNA vaccine can inhibit airway remodeling by regulating the development of Th1/Th2 subsets in asthma mice, and HSP70 may be a potential target for inhaled glucocorticoids (ICS) in the treatment of asthma [128, 129]. Another study also found that HSP70 directly inhibited irritation-induced gastric ulcer formation and promoted gastric ulcer healing [130]. HSP70 can also protect the gastric mucosa through inhibition of apoptosis, proinflammatory cytokines, and cell adhesion molecules [131, 132]. *H. pylori* infection alters gastric epithelial cell proliferation and reduces or even abolishes HSP70 gene expression [133, 134]. The possible mechanisms included inducing the cellular protective effect of HSP70 against *H. pylori* infection via inhibiting the expression of inducible nitric oxide synthase (iNOS). However, the reliability and accuracy, as well as the un-

derlying mechanisms, in this relationship remains poorly understood, and large-sample clinical research must be performed to verify this theory [135, 136]. The direct mechanism of HSP70 related *H. pylori* in protecting asthma remains to be further explored.

The “Disappearing Microbiota” Hypothesis

H. pylori is a kind of ancient, dominant bacteria that settle in the human stomach and closely attach to host cells. *H. pylori* might be a regular member in the gastric microflora in human. With the improvement of environment and lifestyle, and the eradication of *H. pylori*, the prevalence of *H. pylori* infection in the developed countries has declined sharply, while the incidence of asthma, obesity, and allergic diseases has increased rapidly [137–139]. According to Blazer [140], the proponent of the “disappearing microbiota” hypothesis, the absence of certain bacterial species from the ancestral microbiota could

change the environment where cognitive, metabolic, and immunological functions develop in our early life. This change could result in the increased susceptibility to some diseases. The disappearance of ancient microbiota may be a universal paradigm leading to modern diseases. This harmful trend implies that we need to put efforts in understanding and reversing the reasons contributing to the disappearing microbiota [5].

Treatment of Asthma Related to *H. pylori* Infection

Eradicating *H. pylori* could decrease the occurrence of dyspepsia, peptic ulcer, and gastric malignancy. However, concerns of extensive application of eradication treatment are also raised, such as the resistance to antimicrobial agents and an increase in the prevalence of disorders that are negatively correlated with the infection of *H. pylori*, including obesity, asthma, GERD, and Barrett esophagus [141, 142]. Eradicating the infection of *H. pylori* is considered as a double-edged sword. Thus, selective identification and elimination of only the virulent strains of *H. pylori* are of great importance in the eradication therapy [143]. Epidemiology studies and experiments have demonstrated that exposure to *H. pylori* could prevent asthma, especially in one's childhood. Recently, in vivo studies have shown that live bacteria are not involved in induction of this protective role. Administering an extract of *H. pylori* in a newborn could prevent inflammation in the airway and metaplasia of the goblet cells. Injection of *H. pylori* extract could inhibit DCs in processing the allergen in the mediastinal lymph nodes and lungs. These results suggest that the extract of *H. pylori* following sensitization could effectively prevent allergic airway disorders [144]. *H. pylori* targets DCs and relies on highly suppressed Tregs. Since HP-NAP is considered as a possible regulator for Tregs and can inhibit allergic inflammation of asthma, it is possible to develop HP-NAP as an efficient *H. pylori*-specific vaccine to treat allergic asthma [78, 145, 146]. van Wijck et al. [147] have shown that *H. pylori* extract can effectively reduce the production of mucus and multiple characteristics of inflammation in the mice rechallenged by house dust mite. VacA and GGT, 2 persistence determinants in *H. pylori*, are sufficient in preventing asthma and could be given in their purified forms for treatment [148]. Transmaternal *H. pylori* exposure can reduce allergic airway inflammation in the offspring through Tregs and also provide new insights for interventional therapy of asthma [73]. High doses of vitamin D and fish oil supplements during pregnancy have been shown to help prevent

and control disease in the offspring [149], and maternal treatment of Zika virus infection with the IL-1 receptor antagonist can directly reduce fetal neuroinflammatory response through placental immunity [150]. Immunological methods can be used to design vaccines against *H. pylori* infection, and it should also be used for the prevention of asthma across generations [64, 68, 70]. VacA, GGT, HP-NAP, Tregs, and even FOXP3 each play an important role in *H. pylori*-related asthma protection. It would be a very interesting topic if we could design an effective monoepitope or multiepitope vaccine that could be used by the mother before pregnancy, during pregnancy, or during breastfeeding to prevent asthma of the offspring through the placenta or breast milk.

Conclusion

H. pylori may protect allergic asthma by regulating Th1/Th2 and Th17/Tregs balance, inhibiting DCs and HSP70, activating TLRs, and reducing gastroesophageal reflux. The hygiene hypothesis, the "disappearing microbiota" hypothesis, and the gut-lung axis theory all support this protective effect. Therapeutic products made by *H. pylori* may be used to prevent and treat asthma. In particular, perinatal exposure to *H. pylori* can reduce allergic airway inflammation in the offspring, which also provides a new insight for interventional treatment of asthma.

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Conflict of Interest Statement

The authors have no conflicts of interest to declare.

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Author Contributions

Z.Z. and F.Y. conceived and designed this study and contributed equally to this work. Y.M., Y.S., C.B., and C.L. participated actively in the study and approved the submitted manuscript.

References

- 1 Kalach N, Bontems P, Raymond J. Helicobacter pylori infection in children. *Helicobacter*. 2017;22(Suppl 1):e12414.
- 2 Ness-Jensen E, Langhammer A, Hveem K, Lu Y. Helicobacter pylori in relation to asthma and allergy modified by abdominal obesity: the HUNT study in Norway. *World Allergy Organ J*. 2019;12(5):100035.
- 3 Melby KK, Carlsen KL, Håland G, Samdal HH, Carlsen KH. Helicobacter pylori in early childhood and asthma in adolescence. *BMC Res Notes*. 2020;13(1):79.
- 4 Fouda EM, Kamel TB, Nabih ES, Abdelazem AA. Helicobacter pylori seropositivity protects against childhood asthma and inversely correlates to its clinical and functional severity. *Allergol Immunopathol*. 2018;46(1):76–81.
- 5 Tsigalou C, Konstantinidis TG, Cassimos D, Karvelas A, Grapsa A, Tsalkidis A, et al. Inverse association between Helicobacter pylori infection and childhood asthma in Greece: a case-control study. *Germes*. 2019;9(4):182–7.
- 6 Kato H, Pattison R, Bhandari S. Inverse relationship between Helicobacter pylori infection and asthma in US adults with peptic ulcer disease. *Indian J Gastroenterol*. 2017;36(2):158–9.
- 7 Chen C, Xun P, Tsinovoi C, He K. Accumulated evidence on Helicobacter pylori infection and the risk of asthma: a meta-analysis. *Ann Allergy Asthma Immunol*. 2017;119(2):137–45.e2.
- 8 Lim JH, Kim N, Lim SH, Kwon JW, Shin CM, Chang YS, et al. Inverse relationship between Helicobacter pylori infection and asthma among adults younger than 40 years: a cross-sectional study. *Medicine*. 2016;95(8):e2609.
- 9 Iizasa H, Ishihara S, Richardo T, Kanehiro Y, Yoshiyama H. Dysbiotic infection in the stomach. *World J Gastroenterol*. 2015;21(40):11450–7.
- 10 Wang YC, Lin TY, Shang ST, Chen HJ, Kao CH, Wu CC, et al. Helicobacter pylori infection increases the risk of adult-onset asthma: a nationwide cohort study. *Eur J Clin Microbiol Infect Dis*. 2017;36(9):1587–94.
- 11 Molina-Infante J, Gutierrez-Junquera C, Savarino E, Penagini R, Modolell I, Bartolo O, et al. Helicobacter pylori infection does not protect against eosinophilic esophagitis: results from a large multicenter case-control study. *Am J Gastroenterol*. 2018;113(7):972–9.
- 12 Holster IL, Vila AM, Caudri D, den Hoed CM, Perez-Perez GI, Blaser MJ, et al. The impact of Helicobacter pylori on atopic disorders in childhood. *Helicobacter*. 2012;17(3):232–7.
- 13 Wang Q, Yu C, Sun Y. The association between asthma and Helicobacter pylori: a meta-analysis. *Helicobacter*. 2013;18(1):41–53.
- 14 Wang Y, Bi Y, Zhang L, Wang C. Is Helicobacter pylori infection associated with asthma risk? A meta-analysis based on 770 cases and 785 controls. *Int J Med Sci*. 2012;9(7):603–10.
- 15 Chang SS, Hu HY. No inverse relationship between Helicobacter pylori infection and adult asthma with peptic ulcer disease. *Hepatogastroenterology*. 2014;61(130):529–34.
- 16 Sabbagh P, Javanian M, Koppolu V, Vasigala VR, Ebrahimpour S. Helicobacter pylori infection in children: an overview of diagnostic methods. *Eur J Clin Microbiol Infect Dis*. 2019;38(6):1035–45.
- 17 Nakashima S, Kakugawa T, Yura H, Tomonaga M, Harada T, Hara A, et al. Identification of Helicobacter pylori VacA in human lung and its effects on lung cells. *Biochem Biophys Res Commun*. 2015;460(3):721–6.
- 18 Huang YJ, Kim E, Cox MJ, Brodie EL, Brown R, Wiener-Kronish JP, et al. A persistent and diverse airway microbiota present during chronic obstructive pulmonary disease exacerbations. *OMICS*. 2010;14(1):9–59.
- 19 Karimi A, Fakhimi-Derakhshan K, Imanzadeh F, Rezaei M, Cavoshzadeh Z, Maham S. Helicobacter pylori infection and pediatric asthma. *Iran J Microbiol*. 2013;5(2):132–5.
- 20 Taye B, Enquesslassie F, Tsegaye A, Amberbir A, Medhin G, Fogarty A, et al. Association between infection with Helicobacter pylori and atopy in young Ethiopian children: a longitudinal study. *Clin Exp Allergy*. 2017;47(10):1299–308.
- 21 Popescu D, Andronescu D, Babes PA. The association between Helicobacter pylori infection and liver and biliary tract disorders. *Curr Health Sci J*. 2018;44(2):186–91.
- 22 Hossain FMA, Choi JY, Uyangaa E, Park SO, Eo SK. The interplay between host immunity and respiratory viral infection in asthma exacerbation. *Immune Netw*. 2019;19(5):e31.
- 23 Eisenbarth SC, Cassel S, Bottomly K. Understanding asthma pathogenesis: linking innate and adaptive immunity. *Curr Opin Pediatr*. 2004;16(6):659–66.
- 24 Moui A, Klein M, Hassoun D, Dijoux E, Cheminant MA, Magnan A, et al. The IL-15/sIL-15Ra complex modulates immunity without effect on asthma features in mouse. *Respir Res*. 2020;21(1):33.
- 25 Shahir M, Mahmoud Hashemi S, Asadirad A, Varahram M, Kazempour-Dizaji M, Folkerts G, et al. Effect of mesenchymal stem cell-derived exosomes on the induction of mouse tolerogenic dendritic cells. *J Cell Physiol*. 2020;235:7043–55.
- 26 Arikoglu T, Akyilmaz E, Yildirim DD, Batmaz SB, Ulger ST, Aslan G, et al. The relation of innate and adaptive immunity with viral-induced acute asthma attacks: focusing on IP-10 and cathelicidin. *Allergol Immunopathol*. 2017;45(2):160–8.
- 27 Lin CL, Hsiao G, Wang CC, Lee YL. Imperatorin exerts antiallergic effects in Th2-mediated allergic asthma via induction of IL-10-producing regulatory T cells by modulating the function of dendritic cells. *Pharmacol Res*. 2016;110:111–21.
- 28 Abdelaziz MH, Abdelwahab SF, Wan J, Cai W, Huixuan W, Jianjun C, et al. Alternatively activated macrophages; a double-edged sword in allergic asthma. *J Transl Med*. 2020;18(1):58.
- 29 Puggioni F, Alves-Correia M, Mohamed MF, Stomeo N, Mager R, Marinoni M, et al. Immunostimulants in respiratory diseases: focus on Pidotimod. *Multidiscip Respir Med*. 2019;14:31.
- 30 Ege MJ. The hygiene hypothesis in the age of the microbiome. *Ann Am Thorac Soc*. 2017;14(Suppl 5):S348–53.
- 31 Miftahussurur M, Nusi IA, Graham DY, Yamaoka Y. Helicobacter . Hygiene, atopy, and asthma. *Front Microbiol*. 2017;8:1034.
- 32 Jones MG. Understanding of the molecular mechanisms of allergy. *Methods Mol Biol*. 2019;2020:1–15.
- 33 Leaker BR, Singh D, Lindgren S, Almqvist G, Eriksson L, Young B, et al. Effects of the Toll-like receptor 7 (TLR7) agonist, AZD8848, on allergen-induced responses in patients with mild asthma: a double-blind, randomised, parallel-group study. *Respir Res*. 2019;20(1):288.
- 34 Matsushima K, Nagai S. Unraveling the mystery of the hygiene hypothesis through Helicobacter pylori infection. *J Clin Invest*. 2012;122(3):801–4.
- 35 Palomares O, Yaman G, Azkur AK, Akkoc T, Akdis M, Akdis CA. Role of Treg in immune regulation of allergic diseases. *Eur J Immunol*. 2010;40(5):1232–40.
- 36 Daschner A, González Fernández J. Allergy in an evolutionary framework. *J Mol Evol*. 2020;88(1):66–76.
- 37 Asayama K, Kobayashi T, D'Alessandro-Gabazza CN, Toda M, Yasuma T, Fujimoto H, et al. Protein S protects against allergic bronchial asthma by modulating Th1/Th2 balance. *Allergy*. 2020;75(9):2267–78. <https://doi.org/10.1111/all.14261>.
- 38 Hwang YH, Kim SJ, Yee ST. Physcion-matured dendritic cells induce the differentiation of Th1 cells. *Int J Mol Sci*. 2020;21(5):1753.
- 39 Hu C, Li Z, Feng J, Tang Y, Qin L, Hu X, et al. Glucocorticoids modulate Th1 and Th2 responses in asthmatic mouse models by inhibition of notch1 signaling. *Int Arch Allergy Immunol*. 2018;175(1–2):44–52.
- 40 Chen Z, Liu N, Xiao J, Wang Y, Dong R. The amygdala via the paraventricular nucleus regulates asthma attack in rats. *CNS Neurosci Ther*. 2020;26(7):730–40.
- 41 Hwang YH, Paik MJ, Yee ST. Diisononyl phthalate induces asthma via modulation of Th1/Th2 equilibrium. *Toxicol Lett*. 2017;272:49–59.
- 42 Hong ZW, Yang YC, Pan T, Tzeng HF, Fu HW. Differential effects of DEAE negative mode chromatography and gel-filtration chromatography on the charge status of Helicobacter pylori neutrophil-activating protein. *PLoS One*. 2017;12(3):e0173632.

- 43 Amedei A, Codolo G, Del Prete G, de Bernard M, D'Elios MM. The effect of *Helicobacter pylori* on asthma and allergy. *J Asthma Allergy*. 2010;3:139–47.
- 44 D'Elios MM, Codolo G, Amedei A, Mazzi P, Berton G, Zanotti G, et al. *Helicobacter pylori*, asthma and allergy. *FEMS Immunol Med Microbiol*. 2009;56(1):1–8.
- 45 Konturek PC, Rienecker H, Hahn EG, Raithe M. *Helicobacter pylori* as a protective factor against food allergy. *Med Sci Monit*. 2008;14(9):CR452–8.
- 46 Karakullukcu A, Tokman HB, Nepesov S, Demirci M, Saribas S, Vehid S, et al. The protective role of *Helicobacter pylori* neutrophil-activating protein in childhood asthma. *Allergol Immunopathol*. 2017;45(6):521–7.
- 47 Zhou S, Huang Y, Liang B, Dong H, Yao S, Chen Y, et al. Systemic and mucosal pre-administration of recombinant *Helicobacter pylori* neutrophil-activating protein prevents ovalbumin-induced allergic asthma in mice. *FEMS Microbiol Lett*. 2017;364(2):fnw288.
- 48 Liu X, Fu G, Ji Z, Huang X, Ding C, Jiang H, et al. A recombinant DNA plasmid encoding the sIL-4R-NAP fusion protein suppress airway inflammation in an OVA-induced mouse model of asthma. *Inflammation*. 2016;39(4):1434–40.
- 49 Zhao Y, Yang J, Gao YD, Guo W. Th17 immunity in patients with allergic asthma. *Int Arch Allergy Immunol*. 2010;151(4):297–307.
- 50 Massoud AH, Charbonnier LM, Lopez D, Pellegrini M, Phipatanakul W, Chatila TA. An asthma-associated IL4R variant exacerbates airway inflammation by promoting conversion of regulatory T cells to TH17-like cells. *Nat Med*. 2016;22(9):1013–22.
- 51 Wang ZE, Zhou XN, Yang Y, Liu ZY. [Effect of Jian'erle granule on Th17/Treg imbalance of asthma mice]. *Zhongguo Zhong Xi Yi Jie He Za Zhi*. 2016;36(12):1510–4.
- 52 Zhu J, Yamane H, Paul WE. Differentiation of effector CD4 T cell populations. *Annu Rev Immunol*. 2010;28:445–89.
- 53 Peng J, Li XM, Zhang GR, Cheng Y, Chen X, Gu W, et al. TNF-TNFR2 signaling inhibits Th2 and Th17 polarization and alleviates allergic airway inflammation. *Int Arch Allergy Immunol*. 2019;178(3):281–90.
- 54 Park SC, Kim H, Bak Y, Shim D, Kwon KW, Kim CH, et al. An alternative dendritic cell-induced murine model of asthma exhibiting a robust Th2/Th17-skewed response. *Allergy Asthma Immunol Res*. 2020;12(3):537–55.
- 55 Go HN, Lee SH, Cho HJ, Ahn JR, Kang MJ, Lee SY, et al. Effects of chloromethylisothiazolinone/methylisothiazolinone (CMIT/MIT) on Th2/Th17-related immune modulation in an atopic dermatitis mouse model. *Sci Rep*. 2020;10(1):4099.
- 56 Lin CC, Wang YY, Chen SM, Liu YT, Li JQ, Li F, et al. Shegan-Mahuang decoction ameliorates asthmatic airway hyperresponsiveness by downregulating Th2/Th17 cells but upregulating CD4+FoxP3+ Tregs. *J Ethnopharmacol*. 2020;253:112656.
- 57 Lambrecht BN, Hammad H. The immunology of asthma. *Nat Immunol*. 2015;16(1):45–56.
- 58 Choy DF, Hart KM, Borthwick LA, Shikotra A, Nagarkar DR, Siddiqui S, et al. TH2 and TH17 inflammatory pathways are reciprocally regulated in asthma. *Sci Transl Med*. 2015;7(301):301ra129.
- 59 Li HT, Lin YS, Ye QM, Yang XN, Zou XL, Yang HL, et al. Airway inflammation and remodeling of cigarette smoking exposure ovalbumin-induced asthma is alleviated by CpG oligodeoxynucleotides via affecting dendritic cell-mediated Th17 polarization. *Int Immunopharmacol*. 2020;82:106361.
- 60 Nadeem A, Ahmad SF, Al-Harbi NO, Ibrahim KE, Siddiqui N, Al-Harbi MM, et al. Inhibition of Bruton's tyrosine kinase and IL-2 inducible T-cell kinase suppresses both neutrophilic and eosinophilic airway inflammation in a cockroach allergen extract-induced mixed granulocytic mouse model of asthma using preventative and therapeutic strategy. *Pharmacol Res*. 2019;148:104441.
- 61 Santana FPR, da Silva RC, Grecco SDS, Pinheiro AJMCR, Caperuto LC, Arantes-Costa FM, et al. Inhibition of MAPK and STAT3-SOCS3 by sakuranetin attenuated chronic allergic airway inflammation in mice. *Mediators Inflamm*. 2019;2019:1356356.
- 62 Cui H, Huang J, Lu M, Zhang Q, Qin W, Zhao Y, et al. Antagonistic effect of vitamin E on nA2O3-induced exacerbation of Th2 and Th17-mediated allergic asthma via oxidative stress. *Environ Pollut*. 2019;252(Pt B):1519–31.
- 63 Kardan M, Rafiei A, Ghaffari J, Valadan R, Morsaljahani Z, Haj-Ghorbani ST. Effect of ginger extract on expression of GATA3, T-bet and ROR- γ t in peripheral blood mononuclear cells of patients with allergic asthma. *Allergol Immunopathol*. 2019;47(4):378–85.
- 64 Nemattalab M, Shenagari M, Taheri M, Mahjoob M, Nazari Chamaki F, Mojtahedi A, et al. Co-expression of Interleukin-17A molecular adjuvant and prophylactic *Helicobacter pylori* genetic vaccine could cause sterile immunity in Treg suppressed mice. *Cytokine*. 2020;126:154866.
- 65 Lina T, Gonzalez J, Pinchuk I, Beswick E, Reyes V. *Helicobacter pylori* elicits B7H3 expression on gastric epithelial cells: implications in local T cell regulation and subset development during infection. *Clin Oncol Res*. 2019;2(5):2–12.
- 66 Kyburz A, Müller A. *Helicobacter pylori* and extragastric diseases. *Curr Top Microbiol Immunol*. 2017;400:325–47.
- 67 Jafarzadeh A, Larussa T, Nemati M, Jalapour S. T cell subsets play an important role in the determination of the clinical outcome of *Helicobacter pylori* infection. *Microb Pathog*. 2018;116:227–36.
- 68 Lehours P, Ferrero RL. Review: *Helicobacter*: inflammation, immunology, and vaccines. *Helicobacter*. 2019;24(Suppl 1):e12644.
- 69 Gong Y, Tao L, Jing L, Liu D, Hu S, Liu W, et al. Association of TLR4 and Treg in *Helicobacter pylori* colonization and inflammation in mice. *PLoS One*. 2016;11(2):e0149629.
- 70 Nezafat N, Eslami M, Negahdaripour M, Rahbar MR, Ghasemi Y. Designing an efficient multi-epitope oral vaccine against *Helicobacter pylori* using immunoinformatics and structural vaccinology approaches. *Mol Biosyst*. 2017;13(4):699–713.
- 71 Arnold IC, Dehzad N, Reuter S, Martin H, Becher B, Taube C, et al. *Helicobacter pylori* infection prevents allergic asthma in mouse models through the induction of regulatory T cells. *J Clin Invest*. 2011;121(8):3088–93.
- 72 Hussain K, Letley DP, Greenaway AB, Kenefick R, Winter JA, Tomlinson W, et al. *Helicobacter pylori*-mediated protection from allergy is associated with IL-10-secreting peripheral blood regulatory T cells. *Front Immunol*. 2016;7:71.
- 73 Kyburz A, Fallegger A, Zhang X, Altobelli A, Artola-Boran M, Borbet T, et al. Transmaternal *Helicobacter pylori* exposure reduces allergic airway inflammation in offspring through regulatory T cells. *J Allergy Clin Immunol*. 2019;143(4):1496–e11.
- 74 Lambrecht BN, Hammad H. Biology of lung dendritic cells at the origin of asthma. *Immunity*. 2009;31(3):412–24.
- 75 Lee DCP, Tay NQ, Thian M, Prabhu N, Furuhashi K, Kemeny DM. Prior exposure to inhaled allergen enhances anti-viral immunity and T cell priming by dendritic cells. *PLoS One*. 2018;13(1):e0190063.
- 76 Amon L, Lehmann CHK, Baranska A, Schoen J, Heger L, Dudziak D. Transcriptional control of dendritic cell development and functions. *Int Rev Cell Mol Biol*. 2019;349:55–151.
- 77 Oertli M, Müller A. *Helicobacter pylori* targets dendritic cells to induce immune tolerance, promote persistence and confer protection against allergic asthma. *Gut Microbes*. 2012;3(6):566–71.
- 78 Arnold IC, Hitzler I, Müller A. The immunomodulatory properties of *Helicobacter pylori* confer protection against allergic and chronic inflammatory disorders. *Front Cell Infect Microbiol*. 2012;2:10.
- 79 Oertli M, Sundquist M, Hitzler I, Engler DB, Arnold IC, Reuter S, et al. DC-derived IL-18 drives Treg differentiation, murine *Helicobacter pylori*-specific immune tolerance, and asthma protection. *J Clin Invest*. 2012;122(3):1082–96.
- 80 Oertli M, Noben M, Engler DB, Semper RP, Reuter S, Maxeiner J, et al. *Helicobacter pylori* γ -glutamyl transpeptidase and vacuolating cytotoxin promote gastric persistence and immune tolerance. *Proc Natl Acad Sci USA*. 2013;110(8):3047–52.
- 81 Shiu J, Czinn SJ, Kobayashi KS, Sun Y, Blanchard TG. IRAK-M expression limits dendritic cell activation and proinflammatory cytokine production in response to *Helicobacter pylori*. *PLoS One*. 2013;8(6):e66914.

- 82 Farrokhi S, Abbasirad N, Movahed A, Khazaei HA, Pishjoo M, Rezaei N. TLR9-based immunotherapy for the treatment of allergic diseases. *Immunotherapy*. 2017;9(4):339–46.
- 83 Månsson Kvarnhammar A, Tengroth L, Adner M, Cardell LO. Innate immune receptors in human airway smooth muscle cells: activation by TLR1/2, TLR3, TLR4, TLR7 and NOD1 agonists. *PLoS One*. 2013;8(7):e68701.
- 84 Lucas K, Maes M. Role of the Toll like receptor (TLR) radical cycle in chronic inflammation: possible treatments targeting the TLR4 pathway. *Mol Neurobiol*. 2013;48(1):190–204.
- 85 George JA, Park SO, Choi JY, Uyangaa E, Eo SK. Double-faced implication of CD4(+) Foxp3(+) regulatory T cells expanded by acute dengue infection via TLR2/MyD88 pathway. *Eur J Immunol*. 2020;50(7):1000–18.
- 86 Nawijn MC, Motta AC, Gras R, Shirinbak S, Maazi H, van Oosterhout AJ. TLR-2 activation induces regulatory T cells and long-term suppression of asthma manifestations in mice. *PLoS One*. 2013;8(2):e55307.
- 87 Sanchez-Zaucó N, Del Río-Navarro B, Gallardo-Casas C, Del Río-Chivardi J, Muriel-Vizcaino R, Rivera-Pazos C, et al. High expression of Toll-like receptors 2 and 9 and Th1/Th2 cytokines profile in obese asthmatic children. *Allergy Asthma Proc*. 2014;35(3):34–41.
- 88 Piao W, Xiong Y, Li L, Saxena V, Smith KD, Hippen KL, et al. Regulatory T cells condition lymphatic endothelia for enhanced transendothelial migration. *Cell Rep*. 2020;30(4):1052–62.e5.
- 89 Thio CL, Lai AC, Chi PY, Webster G, Chang YJ. Toll-like receptor 9-dependent interferon production prevents group 2 innate lymphoid cell-driven airway hyperreactivity. *J Allergy Clin Immunol*. 2019;144(3):682–97.e9.
- 90 Shalaby KH, Al Heialy S, Tsuchiya K, Farahnak S, McGovern TK, Risse PA, et al. The TLR4-TRIF pathway can protect against the development of experimental allergic asthma. *Immunology*. 2017;152(1):138–49.
- 91 Dong M, Yu H, Wang Y, Sun C, Chang Y, Yin Q, et al. Critical role of toll-like receptor 4 (TLR4) in ricin toxin-induced inflammatory responses in macrophages. *Toxicol Lett*. 2020;321:54–60.
- 92 Borbet TC, Zhang X, Muller A, Blaser MJ. The role of the changing human microbiome in the asthma pandemic. *J Allergy Clin Immunol*. 2019;144(6):1457–66.
- 93 Pachathundikandi SK, Muller A, Backert S. Inflammasome activation by *Helicobacter pylori* and its implications for persistence and immunity. *Curr Top Microbiol Immunol*. 2016;397:117–31.
- 94 Koch KN, Hartung ML, Urban S, Kyburz A, Bahlmann AS, Lind J, et al. *Helicobacter urease*-induced activation of the TLR2/NLRP3/IL-18 axis protects against asthma. *J Clin Invest*. 2015;125(8):3297–302.
- 95 Pachathundikandi SK, Blaser N, Backert S. Mechanisms of inflammasome signaling, microRNA induction and resolution of inflammation by *Helicobacter pylori*. *Curr Top Microbiol Immunol*. 2019;421:267–302.
- 96 Ying L, Ferrero RL. Role of NOD1 and ALPK1/TIFA signalling in innate immunity against *Helicobacter pylori* infection. *Curr Top Microbiol Immunol*. 2019;421:159–77.
- 97 van Heel DA, Ghosh S, Butler M, Hunt K, Foxwell BM, Mengin-Lecreux D, et al. Synergistic enhancement of Toll-like receptor responses by NOD1 activation. *Eur J Immunol*. 2005;35(8):2471–6.
- 98 Ng MT, Van't Hof R, Crockett JC, Hope ME, Berry S, Thomson J, et al. Increase in NF-kappaB binding affinity of the variant C allele of the toll-like receptor 9-1237T/C polymorphism is associated with *Helicobacter pylori*-induced gastric disease. *Infect Immun*. 2010;78(3):1345–52.
- 99 Dela Pena-Ponce MG, Jimenez MT, Hansen LM, Solnick JV, Miller LA. The *Helicobacter pylori* type IV secretion system promotes IL-8 synthesis in a model of pediatric airway epithelium via p38 MAP kinase. *PLoS One*. 2017;12(8):e0183324.
- 100 Kumar P, Singh G, Goyal JP, Khara D, Singh K. Association of common comorbidities with asthma in children: a cross-sectional study. *Sudan J Paediatr*. 2019;19(2):88–92.
- 101 Rogliani P, Sforza M, Calzetta L. The impact of comorbidities on severe asthma. *Curr Opin Pulm Med*. 2020;26(1):47–55.
- 102 Lankarani KB, Honarvar B, Athari SS. The mechanisms underlying *Helicobacter pylori*-mediated protection against allergic asthma. *Tanaffos*. 2017;16(4):251–9.
- 103 Hufnagl K, Pali-Scholl I, Roth-Walter F, Jensen-Jarolim E. Dysbiosis of the gut and lung microbiome has a role in asthma. *Semin Immunopathol*. 2020;42(1):75–93.
- 104 Ver Heul A, Planer J, Kau AL. The human microbiota and asthma. *Clin Rev Allergy Immunol*. 2019;57(3):350–63.
- 105 Lérias JR, Parascoudi G, de Sousa E, Martins J, Condeco C, Figueiredo N, et al. Microbes as master immunomodulators: immunopathology, cancer and personalized immunotherapies. *Front Cell Dev Biol*. 2019;7:362.
- 106 McAleer JP, Kolls JK. Contributions of the intestinal microbiome in lung immunity. *Eur J Immunol*. 2018;48(1):39–49.
- 107 Budden KF, Gellatly SL, Wood DL, Cooper MA, Morrison M, Hugenholtz P, et al. Emerging pathogenic links between microbiota and the gut-lung axis. *Nat Rev Microbiol*. 2017;15(1):55–63.
- 108 Bassis CM, Erb-Downward JR, Dickson RP, Freeman CM, Schmidt TM, Young VB, et al. Analysis of the upper respiratory tract microbiotas as the source of the lung and gastric microbiotas in healthy individuals. *mBio*. 2015;6(2):e00037.
- 109 Mukherjee S, Hanidziar D. More of the gut in the lung: how two microbiomes meet in ARDS. *Yale J Biol Med*. 2018;91(2):143–9.
- 110 He Y, Wen Q, Yao F, Xu D, Huang Y, Wang J. Gut-lung axis: the microbial contributions and clinical implications. *Crit Rev Microbiol*. 2017;43(1):81–95.
- 111 Anand S, Mande SS. Diet, microbiota and gut-lung connection. *Front Microbiol*. 2018;9:2147.
- 112 Frati F, Salvatori C, Incorvaia C, Bellucci A, Di Cara G, Marcucci F, et al. The role of the microbiome in asthma: the gut(-)lung axis. *Int J Mol Sci*. 2018;20(1):123.
- 113 Samuelson DR, Welsh DA, Shellito JE. Regulation of lung immunity and host defense by the intestinal microbiota. *Front Microbiol*. 2015;6:1085.
- 114 Gupta N, Kumar R, Agrawal B. New players in immunity to tuberculosis: the host microbiome, lung epithelium, and innate immune cells. *Front Immunol*. 2018;9:709.
- 115 Wang J, Li F, Wei H, Lian ZX, Sun R, Tian Z. Respiratory influenza virus infection induces intestinal immune injury via microbiota-mediated Th17 cell-dependent inflammation. *J Exp Med*. 2014;211(12):2397–410.
- 116 Vital M, Harkema JR, Rizzo M, Tiedje J, Brandenberger C. Alterations of the murine gut microbiome with age and allergic airway disease. *J Immunol Res*. 2015;2015:892568.
- 117 Ojha UC, Singh DP, Choudhari OK, Gothi D, Singh S. Correlation of severity of functional gastrointestinal disease symptoms with that of asthma and chronic obstructive pulmonary disease: a multicenter study. *Int J Appl Basic Med Res*. 2018;8(2):83–8.
- 118 Hauptmann M, Schaible UE. Linking microbiota and respiratory disease. *FEBS Lett*. 2016;590(21):3721–38.
- 119 Stiemsma LT, Turvey SE. Asthma and the microbiome: defining the critical window in early life. *Allergy Asthma Clin Immunol*. 2017;13:3.
- 120 Mendez R, Banerjee S, Bhattacharya SK, Banerjee S. Lung inflammation and disease: a perspective on microbial homeostasis and metabolism. *IUBMB Life*. 2019;71(2):152–65.
- 121 Cheng M, Cao L, Ning K. Microbiome big-data mining and applications using single-cell technologies and metagenomics approaches toward precision medicine. *Front Genet*. 2019;10:972.
- 122 Chiu L, Bazin T, Truchetet ME, Schaeverbeke T, Delhaes L, Pradeu T. Protective microbiota: from localized to long-reaching co-immunity. *Front Immunol*. 2017;8:1678.
- 123 Baturcam E, Snape N, Yeo TH, Schagen J, Thomas E, Logan J, et al. Human metapneumovirus impairs apoptosis of nasal epithelial cells in asthma via HSP70. *J Innate Immun*. 2017;9(1):52–64.

- 124 Liu L, Huang Y, Feng X, Chen J, Duan Y. Overexpressed Hsp70 alleviated formaldehyde-induced apoptosis partly via PI3K/Akt signaling pathway in human bronchial epithelial cells. *Environ Toxicol*. 2019;34(4):495–504.
- 125 Harkins MS, Moseley PL, Iwamoto GK. Regulation of CD23 in the chronic inflammatory response in asthma: a role for interferon-gamma and heat shock protein 70 in the TH2 environment. *Ann Allergy Asthma Immunol*. 2003;91(6):567–74.
- 126 Yombo DJK, Mentink-Kane MM, Wilson MS, Wynn TA, Madala SK. Heat shock protein 70 is a positive regulator of airway inflammation and goblet cell hyperplasia in a mouse model of allergic airway inflammation. *J Biol Chem*. 2019;294(41):15082–94.
- 127 Huang X, Tan X, Liang Y, Hou C, Qu D, Li M, et al. Differential DAMP release was observed in the sputum of COPD, asthma and asthma-COPD overlap (ACO) patients. *Sci Rep*. 2019;9(1):19241.
- 128 Yan L, Xiao-Ling S, Zheng-Yan C, Guo-Ping L, Sen Z, Zhuang C. HSP70/CD80 DNA vaccine inhibits airway remodeling by regulating the transcription factors T-bet and GATA-3 in a murine model of chronic asthma. *Arch Med Sci*. 2013;9(5):906–15.
- 129 Jiang H, Zhang X, Chi X, Wang J, Wang J, Dou J. [The effect of inhaled glucocorticoid therapy on serum proteomics of asthmatic patients]. *Zhonghua Jie He He Hu Xi Za Zhi*. 2014;37(4):274–8.
- 130 Saremi K, Rad SK, Khalilzadeh M, Hussaini J, Majid NA. In vivo acute toxicity and anti-gastric evaluation of a novel dichloro Schiff base: Bax and HSP70 alteration. *Acta Biochim Biophys Sin*. 2020;52(1):26–37.
- 131 deFonesca A, Kaunitz JD. Gastrointestinal mucosal defense. *Curr Opin Gastroenterol*. 2010;26(6):604–10.
- 132 Sidahmed HM, Azizan AH, Mohan S, Abdulla MA, Abdelwahab SI, Taha MM, et al. Gastroprotective effect of desmosmumotin C isolated from *Mitrella kentii* against ethanol-induced gastric mucosal hemorrhage in rats: possible involvement of glutathione, heat-shock protein-70, sulfhydryl compounds, nitric oxide, and anti-Helicobacter pylori activity. *BMC Complement Altern Med*. 2013;13:183.
- 133 Tao L, Zou H, Huang Z. Effects of Helicobacter pylori and heat shock protein 70 on the proliferation of human gastric epithelial cells. *Gastroenterol Res Pract*. 2014;2014:794342.
- 134 Pierzchalski P, Krawiec A, Ptak-Belowska A, Baranska A, Konturek SJ, Pawlik WW. The mechanism of heat-shock protein 70 gene expression abolition in gastric epithelium caused by Helicobacter pylori infection. *Helicobacter*. 2006;11(2):96–104.
- 135 Qu B, Jia Y, Liu Y, Wang H, Ren G, Wang H. The detection and role of heat shock protein 70 in various nondisease conditions and disease conditions: a literature review. *Cell Stress Chaperones*. 2015;20(6):885–92.
- 136 Targosz A, Pierzchalski P, Krawiec A, Szczyrk U, Brzozowski T, Konturek SJ, et al. Helicobacter pylori inhibits expression of heat shock protein 70 (HSP70) in human epithelial cell line. Importance of Cag A protein. *J Physiol Pharmacol*. 2006;57(2):265–78.
- 137 Taube C, Muller A. The role of Helicobacter pylori infection in the development of allergic asthma. *Expert Rev Respir Med*. 2012;6(4):441–9.
- 138 Otero LL, Ruiz VE, Perez Perez GI. Helicobacter pylori: the balance between a role as colonizer and pathogen. *Best Pract Res Clin Gastroenterol*. 2014;28(6):1017–29.
- 139 Blaser MJ. The Jeremiah Metzger Lecture: global warming redux: the disappearing microbiota and epidemic obesity. *Trans Am Clin Climatol Assoc*. 2012;123:230–8.
- 140 Blaser MJ. The theory of disappearing microbiota and the epidemics of chronic diseases. *Nat Rev Immunol*. 2017;17(8):461–3.
- 141 O'Connor A, O'Morain CA, Ford AC. Population screening and treatment of Helicobacter pylori infection. *Nat Rev Gastroenterol Hepatol*. 2017;14(4):230–40.
- 142 Yap TW, Leow AH, Azmi AN, Callahan DL, Perez-Perez GI, Loke MF, et al. Global fecal and plasma metabolic dynamics related to Helicobacter pylori eradication. *Front Microbiol*. 2017;8:536.
- 143 Abadi AT, Kusters JG. Management of Helicobacter pylori infections. *BMC Gastroenterol*. 2016;16(1):94.
- 144 van Wijck Y, de Kleijn S, John-Schuster G, Mertens TCJ, Hiemstra PS, Muller A, et al. Therapeutic application of an extract of Helicobacter pylori ameliorates the development of allergic airway disease. *J Immunol*. 2018;200(5):1570–9.
- 145 Sehrawat A, Sinha S, Saxena A. Helicobacter pylori neutrophil-activating protein: a potential Treg modulator suppressing allergic asthma? *Front Microbiol*. 2015;6:493.
- 146 D'Elia MM, Andersen LP. Inflammation, immunity, and vaccines for Helicobacter pylori. *Helicobacter*. 2009;14(Suppl 1):21–8.
- 147 van Wijck Y, John-Schuster G, van Schadewijk A, van den Oever RL, Obieglo K, Hiemstra PS, et al. Extract of Helicobacter pylori ameliorates parameters of airway inflammation and goblet cell hyperplasia following repeated allergen exposure. *Int Arch Allergy Immunol*. 2019;180(1):1–9.
- 148 Engler DB, Reuter S, van Wijck Y, Urban S, Kyburz A, Maxeiner J, et al. Effective treatment of allergic airway inflammation with Helicobacter pylori immunomodulators requires BATF3-dependent dendritic cells and IL-10. *Proc Natl Acad Sci U S A*. 2014;111(32):11810–5.
- 149 Bisgaard H, Vissing NH, Carson CG, Bischoff AL, Folsgaard NV, Kreiner-Moller E, et al. Deep phenotyping of the unselected COPSAC2010 birth cohort study. *Clin Exp Allergy*. 2013;43(12):1384–94.
- 150 Lei J, Vermillion MS, Jia B, Xie H, Xie L, McLane MW, et al. IL-1 receptor antagonist therapy mitigates placental dysfunction and perinatal injury following Zika virus infection. *JCI Insight*. 2019;4(7):e122678.