

Sexual Dimorphism in the Expression of Pain Phenotype in Preclinical Models of Rheumatoid Arthritis

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KEYWORDS

• Rheumatoid arthritis • Pain • Sexual dimorphism

KEY POINTS

- Rheumatoid arthritis is the most frequent rheumatic disease with a higher prevalence in females than in males.
- Pain is a cardinal symptom of rheumatoid arthritis and strongly impacts patient quality of life.
- Sexual dimorphism in pain processing has been described in the literature since 1988.
- Sexual dimorphism in rheumatoid arthritis has been reported. However, there remains a dearth of studies directly addressing sexual dimorphism in rheumatoid arthritis pain mechanisms.

A BRIEF OVERVIEW OF RHEUMATOID ARTHRITIS

Historically, rheumatoid arthritis (RA) was first described in [1](#page-10-0)800.¹ It is at present the most frequent chronic inflammatory rheumatic disease.^{[2](#page-10-1)} RA prevalence ranges from 0.3% to 1.0% of the population in industrialized countries according to the World Health Organization^{[3](#page-10-2)[,4](#page-10-3)} and its incidence increases with the aging of the population.^{[5,](#page-10-4)[6](#page-10-5)} This disease is characterized as an autoimmune-mediated inflammatory disease that involves both immunologic activation and inflammatory pathways. Once these pathways are triggered, it results in a self-perpetuating process that leads to joint inflam-mation, cartilage degradation, bone erosion, and chronic pain.^{[7](#page-10-6)} Identification and prognosis of patients with RA have been improved with the recognition of specific anticitrullinated peptide antibodies (ACPA) 8 in addition to rheumatoid factor. 9 Indeed, the detection of anticyclic citrullinated peptides antibodies (anti-CCP) presents with a

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98% specificity and are thus considered as useful tools for the diagnosis of rheuma-toid disease.^{[10](#page-10-9)} Although the dynamic process of rheumatic disease initiation and progression is not clear, a robust association between genetic, $11-15$ epigenetic, $16-21$ environmental, $22-29$ and immunologic $30-34$ components during the development of RA has been demonstrated.

From the genetic perspective, it is well-recognized that RA susceptibility may be strongly linked to family history. 35 The increased prevalence of disease within racial groups has been shown, as in native Americans where prevalence rates of RA of 5% to 7% have been reported.^{[36](#page-12-1)} Since the early 1970s, class II human leukocyte antigens (HLA) were associated with susceptibility to RA. $37,38$ $37,38$ The major HLA susceptible locus associated with RA is *HLA-DRB1*, especially with regard to a specific sequence of amino acids referred to as "the shared epitope" in different *HLA-DRB1* alleles (eg, **01, *04, *10, *14*)[.39](#page-12-4) Specific amino acids at position 11 (eg, Val, Leu, Pro) or 13 (eg, His and Phe) are strongly associated with RA development.^{[40](#page-12-5)[,41](#page-12-6)} Other non-HLA genes have been shown to be associated risk factors for developing RA, including *PTPN22,* TNF, IL-1B, CRP, and CTL4.^{13-15[,42](#page-12-7)} Although these genetic associations with RA susceptibility are robust, there is not a perfect genetic consonance and other factors clearly can contribute to the development of RA. Associated variables that have been strongly implicated in the development of RA include environmental factors, such as cigarette smoking, 22 pollutants (eg, silica), 43 and patient variables (eg, diet), 44 44 44 mucosal microbiomes, $^{28,29,45-47}$ $^{28,29,45-47}$ $^{28,29,45-47}$ $^{28,29,45-47}$ and sex/gender. 48,49 48,49 48,49 48,49

Of particular interest in this review is the issue of the sex linkages and the development and progression of RA. The RA diagnosis in fact presents as a heterogeneous phenotype, the severity and patterns of which are influenced by sex-associated factors. RA is 2 to 5 times more common in women than men, depending on the region in the world.^{5,[6](#page-10-5)} In women, RA onset is commonly seen between 30 to 60 years of age, whereas in men, RA often begins later in life.^{[50](#page-13-2)} Thus, after the age of 65, the ratio of the incidence in men and women shifts with the predominance occurring in men more than 75 years of age.^{[49](#page-13-1)} Although it is not clear which factors drive the sexual dimorphism in patients with RA, comparing clinical data in humans, and preclinical studies in rodents, several hypotheses have been proposed based on sex hormones $51-53$ and immune cells. 54

PAIN IN RHEUMATOID ARTHRITIS Model of Phases of Pain in Rheumatoid Arthritis

The mechanisms underlying pain state in the patient with RA represent complex processes that differ between stages of the disease $(Fig. 1)$ $(Fig. 1)$ $(Fig. 1)$.^{[55](#page-13-5)} The production of autoantibodies (eg, rheumatoid factor, ACPA, anti-CarP) starts months to years before the onset of the disease, gradually increasing^{[56](#page-13-6)} (dark blue line; see [Fig. 1](#page-2-0), \bullet). We now recognize in the early preclinical arthritis phase, patients can suffer from arthralgia (purple line; see [Fig. 1](#page-2-0), \bullet) before the development of clinically evident signs, such as palpable synovitis (dotted blue line; see Fig. $1, 0$). In the subsequent development of active disease (eg, with clinical signs of joint pathology), patients with RA develop chronic inflammatory joint pain, which display a cyclic waxing and waning (see [Fig. 1](#page-2-0), ➋). These clinical signs may then display some degree of remission after treatment with modern therapies. In remission, pain resolves in some patients (see [Fig. 1](#page-2-0), Θ), but in a significant proportion of patients arthralgia persists^{[57](#page-13-7)} (see [Fig. 1](#page-2-0), \odot). Moreover, remission is not necessarily associated with declining autoantibody titers and disappearance of autoantibodies remains rare^{[58](#page-13-8)} (dotted dark blue line; see [Fig. 1](#page-2-0), ➍). Mechanisms involved in RA pain are also different between stages (for an extensive literature on this subject see reviews $55,59-62$ $55,59-62$).

Fig. 1. Pain evolution according to RA disease activity. In RA, pain symptom in patients follows different steps⁵⁵: \bullet in a preclinical stage, patient can develop arthralgia before the onset of the disease potentially associated with an early production of autoantibodies, [●] after RA diagnostic, in clinical stage, marked by the development of synovitis, pain and disease activity both increase and fluctuate without being entirely correlated each other, [●] following an effective therapy, pain symptom can decrease in a remission stage, but \odot in a significant proportion of patients, pain symptoms persist even though the disease activity decreased.

Simplistically, pain processing begins with the activation of nociceptive primary afferents, which innervate the joint. 63 In turn, these nociceptive afferents lead to the activation of second-order neurons in the spinal dorsal horn that project via the ventrolateral long tract to a variety of supraspinal centers in the brain. Some of these projections through the somatosensory thalamus to the somatosensory cortex mediate the so-called sensory discriminative aspect of pain, whereas other projections are through more medial thalamus regions and project to the areas of the limbic forebrain associated with emotion and affect. The consequence is the pain state, which comprises the emotional and affective factors that lead, ultimately, to a protective response. $63,64$ $63,64$ Although the acute activation of these circuits normally have a protective role (eg, to evoke for example limb withdrawal), these states, when chronic, become maladaptive. Such chronic pain attributes in RA have a marked prevalence in females. 65-67 However, there is no common pathophysiologic sex-related mechanism shared by all chronic pain conditions; each one can present specific factors that differ between females and males. 68

Lack of Covariance with Physical Signs and Pain in Rheumatoid Arthritis

An important point to be made in the conceptual summary displayed in [Fig. 1](#page-2-0) is that, although joint morphology, pathology, and loss of function primarily underlie the RA diagnosis, pain displays deviations from the expression of the underlying clinically evident pathology for the patient with RA. Of note, this dissociation has particular relevance in that pain is the cardinal symptom of RA that strongly impacts these patients' quality of life.^{[69](#page-14-1)} In fact, 75% to 80% of patients with RA in treatment have moderate to severe pain symptoms. $70,71$ $70,71$ Only 26% of patients are satisfied with their RA treatment⁶⁹ and between 55% and 65% of patients are unsatisfied with their pain management, 70 a priority for patients with RA. 71 It is important to take into account that most of data are obtained from women (75%–90% depending on the study^{[69,](#page-14-1)[70](#page-14-2)}). The actual "treat to target" strategy for RA aims to decrease the pathologic changes^{[72](#page-14-4)} and thereby to modulate pain. However, joint pain is in fact poorly correlated with the inflammatory state of the patient with RA. 62 62 62 Even with optimal regulation of inflammatory cascades, pain has often been shown to be insufficiently controlled.^{[57](#page-13-7)} It is wellappreciated that joint pain (arthralgia) often begins before other manifestations of joint inflammation $73,74$ $73,74$ and consequently before any diagnosis or the implementation of an RA therapeutic strategy. A further complexity is that, although the RA phenotype involves joint structures (synovia, cartilage, and bones), an autoimmune response outside of the joint is also observed and is usually associated with other pathologies, such as rheumatoid nodules, lymphatic vessel tumefaction, pleuritis, and cardiovas-cular or ocular manifestations.^{[75](#page-14-7)} These observations have led to an appreciation that pain arising in association with RA may reflect parallel processes that may be influenced by the variables that are associated with the physical manifestation of RA and particularly given the role played by sex.

Covariance of Antibodies with Pain and Dissociation from Joint Manifestations

As presented in conceptual summary in [Fig. 1](#page-2-0), arthralgia can in fact be reported months to years before the actual onset of clinically identified disease. It is now appreciated that this onset of arthralgia may occur concurrently with the expression of autoantibodies (ie, rheumatoid factor, 76 ACPA, $77-79$ anti-CarP, $80-82$ and anti-MAA $83,84$ $83,84$). These autoantibodies can be detectable in patients reporting predisease arthralgias and may be associated with persistent arthralgia during remission. Recently, a direct link between ACPA isolated from patients with RA and pain development in mice has been described^{[85](#page-15-0)} and was felt to mechanistically involve chemokine production by os-teoclasts.^{[86](#page-15-1)} Phenotypic changes of in the Fc portion of antibodies, such as galactosylation, fucosylation, or sialylation, could be involved in the proinflammatory profile acquisition and certainly pronociceptive. $87-90$ In addition to autoantibodies, RA is marked by an early alteration of cytokines and chemokines levels, such as IL-5 or IL-17A, $90-93$ which are directly able to sensitize the nociceptor. 63

As indicated in [Fig. 1](#page-2-0), during the clinical stage, synovitis development induces a chronic joint pain marked by peripheral and central sensitization initially induced by immune cells in $RA⁵⁵$ It is interesting to note that a comorbid condition, fibromyalgia, reflects a chronic widespread pain associated with tenderness, sleep disturbance, and psychiatric distress in the absence of clinical signs of inflammation. This condition affects around 14% to 20% of patients with RA, with a higher prevalence in women $.94,95$ $.94,95$

PRECLINICAL MODELS AND SEXUAL DIMORPHISM IN THE PAIN PHENOTYPE OF RHEUMATOID ARTHRITIS

Given the complexities of pain in different stages of arthritis, research efforts have focused on drawing parallels between sex differences observed in RA pathophysiology and sex differences underlying the pain phenotype observed clinically and in preclinical models of RA.

Use of Rodent Models in Arthritis Studies to Study the Role of Sex

Pioneering studies on the sexual dimorphism of pain expression began to emerge around 1988, when Bodnar and associates $96,97$ $96,97$ demonstrated sex differences in basal nociceptive thresholds in rodents that seemed to be modulated by sex hormones. In particular, a large number of publications came in the mid-1990s and drew

considerable attention to the topic. $66,68,98$ $66,68,98$ $66,68,98$ $66,68,98$ Previously, most preclinical studies traditionally used males as subjects and the sex-related differences in pain and their mechanisms were not explored widely. In 2016, the National Institutes of Health initiated a policy requiring preclinical research to use males and females in supported research. Currently, it is clear that males and females do not manifest the same pain experience, showing different physiologic and behaviorally defined pain responses. $67-74$ As reviewed elsewhere in this article, overall, females, in preclinical models, often present a lower threshold and or a heightened pain response than that observed in males. $99-101$

Preclinical animal models of RA have been used to understand the pain phenotype and response to analgesic drugs during development and maintenance of RA and can be used to concurrently examine male and female rodents. Here we introduce animal models of RA that have been used to understand sexual dimorphism in the RA pain phenotype and studies directly addressing different hypotheses in relation to pain mechanisms at the levels of primary afferent neurons, dorsal root ganglia (DRG) and trigeminal ganglia, the dorsal horn of the spinal cord, and supraspinal structures.

Several preclinical models have been widely used to understand pain processing and sexual dimorphism in RA (for review see $61,102,103$ $61,102,103$ $61,102,103$). Broadly speaking, these models represent the induction of monoarthritis or polyarthritis with time-dependent changes in the joint morphology (eg, synovial lesions, bone resorption, and cartilage destruction), in the inflammatory phenotype (eg, synovial inflammation, infiltration of immune cells), and in the development of pronounced and ongoing pain and changes in behavior, including pain evoked behaviors (eg, mechanical and thermal thresholds), pain-suppressed behaviors (eg, feeding, mating, and locomotor activity), and operant conditioning (eg, conditioned place preference).^{[102](#page-15-10)} As will be noted, in comparison with the immunization model, collagen-induced arthritis, the collagen-antibody– induced arthritis (CAIA), and K/BxN models present over time evidence of evolving pain phenotypes.^{[104](#page-16-0)} A brief description of these models is presented.

- i. The complete Freud's adjuvant (CFA) generated by an intradermal injection 105 (pol-yarthritis) or an intra-articular injection of CFA (monoarthritis).^{[106](#page-16-2)[,107](#page-16-3)} This model is mediated by the innate immune system and leads to the infiltration of inflammatory cells, synovial hypertrophy, and joint alterations, as well as inflammatory response in the viscera, skin, and muscle. These rodents develop hypersensitivity to innocuous stimuli with an immediate onset (<24 hours) and persisting for weeks.
- ii. The collagen-induced arthritis model, which is induced by intradermal injection(s) of type II collagen that activate the adaptive immune system and the production of anticollagen II antibodies. This model of polyarthritis leads to a breach of self-tolerance, T- and B-cell activity and activation of anticartilage immunity.^{[108](#page-16-4)} In this model, animals develop hypersensitivity at the onset of the disease (within the second week) that persists for at least 28 days.
- iii. The CAIA model is induced by an intraperitoneal or intravenous injection of a cocktail of anticollagen II antibodies followed by an intraperitoneal injection of lipopolysaccharide to synchronize the onset of inflammation. The CAIA model of polyarthritis does not involve T and B cells, but directly uses antibodies against joint-specific epitopes, leading to synovial inflammation, the infiltration of immune cells, and the destruction of bone and cartilage. In this model, mechanical hypersensitivity precedes the inflammatory phase and persists for 3 to 4 months, even after the resolution of inflammation.^{[61](#page-13-16)}
- iv. The K/BxN passive serum transfer model of polyarthritis is induced by the intraperitoneal injection(s) of sera from KRN-NOD (K/BxN) transgenic mice that

demonstrate an inflammatory arthritis phenotype that begins shortly after weaning and continues unabated into adulthood.^{[109](#page-16-5)} The serum from the transgenic K/BxN mice contains antibodies directed against glucose-6-phosphate isomerase and associated immune complexes, leading to synovial inflammation, the infiltration of immune cells, and the destruction of bone and cartilage. This model is characterized by a biphasic period where tactile and cold allodynia are observed very early after the delivery of the K/BxN serum.^{[110](#page-16-6)} The clinical signs (eg, joint swelling, erythema) resolve and, of note, the allodynia persists unabated. As noted in [Table 1](#page-6-0), the early phase displays an inflammatory analgesic pharmacology, whereas the late phase seems to represent an analgesic pharmacology of a neuropathic state.

As shown in [Table 1](#page-6-0), in the models as described elsewhere in this article, a sexual dimorphism for arthritis score, pain phenotype, neuraxial changes, and response to treatment has been a focus of limited research to date with direct comparisons. Regarding inflammation, females exhibited a higher arthritis score in CFA rats and female rats with collagen-induced arthritis had a greater susceptibility than males, whereas in the CAIA and K/BxN passive transfer models, no difference was observed in arthritis development between sexes (in mice). For the pain phenotype, a sexual dimorphism was observed in female CFA rats that presented a lower evoked paw pressure threshold, whereas there were no differences in the collagen-induced arthritis male and female rats. In contrast, female K/BxN passive serum transfer mice showed a resolution in both paw inflammation and evoked mechanical allodynia pain phenotype in the late phase, whereas males presented a postinflammatory and long-lasting allodynia.

Major sexual dimorphism was observed in the molecular mechanism of pain processing; indeed, in the CAIA and K/BxN models. Female mice showed a spinal microglia (ie, the ionized calcium binding adaptor molecule 1 marker) activation in early and late phases and astrocyte activation (ie, the glial fibrillary acidic protein marker) only in the late phase. Interestingly, K/BxN male mice presented only microglial activation in the late phase and no significant astrocyte activation. In the CAIA model, both males and females showed an increase of spinal calcitonin gene-related peptide and substance P, key neuropeptides involved in central sensitization playing a role in the development and maintenance of hyperalgesia,^{[114](#page-16-7)} but spinal galanin, a neuropeptide known to be involved in neuropathic pain, was only increased in males in the late phase of CAIA. In the K/BxN model, both males and females show an enhanced expression of tumor necrosis factor in the spinal cord at the peak of paw inflammation. However, also in the spinal cord females showed enhanced IL-10 messenger RNA (mRNA) and in the late phase increased type I interferon-beta mRNA expression compared with male mice. These protein and mRNA markers are leading to our cellular and molecular understanding of the sex differences seen in the pain phenotypes in arthritis.

In addition to the differences in the biomarkers of pain, there were sex differences in the responses to pharmacologic treatments. In the CFA model, opioid agonists (eg, morphine, butorphanol, oxycodone, and loperamide) were more potent in males than in females. Intrathecal administration of glial inhibitors using minocycline or pentoxifylline were only effective in male CIAI mice, highlighting a prominent role for microglia in male pain processing. Intrathecal injections also demonstrated that pain modulation was largely at the spinal cord/DRG level in the K/BxN model. In the K/ BxN mice, intrathecal injection of anti-tumor necrosis factor antibodies transiently reduced pain as did the injections of interferon-ß. Neither single agent was effective

Abbreviations: º, female; ♂, male; CFA, complete Freud adjuvant; CIA, collagen-induced arthritis; CAIA, collagen antibody-induced arthritis; K/BxN, K/BxN serum transfer; GFP, glial fibrillary acidic protein; TNF, tumor necrosis factor.

in the long term; however, 2 injections with both agents permanently reversed the pain phenotype in male mice. This result suggested that the spontaneous biologic advantage of self-remitting pain in female mice could be phenocopied with therapeutic injections of biologic agents at the anatomic level of the difference between male and female pain mechanisms (in this case the intrathecal space).

The Role of Sex Hormones

Sexual dimorphism in the pain experience is probably mostly defined by sex hormones. Although the effect of these hormones on pain is complex and not yet completely understood, it is well-known that sex hormones influence both peripheral and central nervous system pathways by modulating neuronal activity.[115](#page-16-12) Considering sex hormones in patients with RA, it is well-known that, during pregnancy, circulating steroid hormones (ie, estrogen, progesterone, and cortisol) seem to directly or indirectly suppress synovial inflammation inhibiting the maternal immune system and inducing an immune tolerance.^{[51](#page-13-3)} Moreover, postpartum, breastfeeding is associated with a lower risk of RA^{52} RA^{52} RA^{52} and the production of prolactin that has immunomodulatory properties.^{[116](#page-16-13)} Furthermore, RA is more frequently observed after menopause and menopause at a young age (\leq 45 years old) is a risk factor for RA.^{[53](#page-13-19)} Some studies also demonstrated a protective effect of progesterone receptor signaling in mice 117 and androgen may also protect against RA development in humans.^{[118](#page-16-15)} Conversely, androgen deprivation therapy^{[119](#page-16-16)} or hypogonadism^{[120](#page-16-17)} increases the risk of RA in men. Therefore, a sexual dimorphism in the expression of pain phenotype in RA is undoubtedly associated, in part, by sex hormones.

Hormonal Receptors on Neurons

Androgen receptors (AR) are expressed in DRG sensory neurons and one-half of ARpositive neurons were nociceptive afferents¹²¹; some studies have demonstrated the involvement of testosterone in pain modulation.^{[122–125](#page-17-0)} It has been shown that primary afferents also express estrogen receptors (ER α and ER β).^{[126,](#page-17-1)[127](#page-17-2)} Preclinical models have shown that $ER\alpha$ expression seems to be restricted to nociceptive afferents whereas $ER\beta$ is more widely distributed.^{[128](#page-17-3)} It has been demonstrated that ERs can play a role in the control of peripheral nociceptive signaling by interacting with purinergic, P2X3, and transient receptor potential vanilloide 1 receptors in nociceptive afferent neurons.^{[127,](#page-17-2)[129–133](#page-17-4)} The expression of both P2X3 and transient receptor potential vanilloide 1 receptors was significantly reduced in female ERs (ER α and ER β) knockout mice compared with the expression in female wild type mice. Ma and col-leagues^{[130](#page-17-5)} demonstrated that ovariectomized female rats had an increase of P2X3 expression in DRG neurons followed by a higher mechanical sensitivity, which were reversed by estrogen replacement. In contrast, mechanical sensitivity and P2X3 expression did not change in the DRG of orchiectomized male rats, evidencing the relation between estrogen and P2X3 expression in nociceptive afferents. Because P2X3 receptors are expressed in the fibers innervating joints, this relation between estrogen and P2X3 receptors might explain, at least in part, the sex differences in pain observed in RA.

In addition, estrogen can also mediate the sex-related differences in substance P release^{[134](#page-17-6)} and in the expression of both nerve growth factor and its high-affinity receptor tropomyosin receptor kinase A in DRG neurons.¹³⁵⁻¹³⁷ Substance P expres-sion is altered in the joint and dorsal horn of animals with arthritis.^{[138,](#page-18-0)[139](#page-18-1)} Nerve growth factor and its receptor are involved in pain^{[140](#page-18-2)} and rheumatic diseases.^{[141](#page-18-3)} Calcitonin gene-related peptide expression in DRG neurons also undergoes an estrogen influence.¹⁴² Therefore, the effects of estrogen on nociceptive neurons may also be contributing to the sexual dimorphism in the pain phenotype of RA. However, studies are needed comparing the female and male pain phenotypes and their nociceptive afferent proteomic profiles. This need is more pronounced when a specific condition is considered. Sensory afferent neurons also express prolactin receptors, which modulate neuronal activity majorly in females.^{[143](#page-18-5)} Patil and colleagues¹⁴³ have shown that prolactin receptor mRNA was expressed equally in female and male peptidergic nociceptors and central terminals; however, prolactin protein was found only in females, and, in turn, prolactin-induced excitability was detected only in female DRG neurons. Strikingly, in patients with RA, both males and females, increased serum levels of prolactin cooperates with other proinflammatory stimuli to activate macrophages, 144 but the clinical significance of prolactin to RA pathogenesis remains unclear. Taking into account these recent data in the literature, it is plausible that, during RA development, primary afferent neurons could play a role in sexual dimorphism of the RA pain phenotype.

Sex Differences in Transcription at the Dorsal Root Ganglion

The sexual dimorphism in response to a noxious stimulus can involve differences at the level of primary afferent neurons, dorsal root and trigeminal ganglia, dorsal horn of the spinal cord and supraspinal structures. DRG and trigeminal ganglia primary afferent neurons have been found as possible source of mechanistic diversity that causes sex differences in pain. $143,145$ $143,145$ In fact, studies suggested sex-related differences in the sensory neuron transcriptome. A transcriptomic analysis of DRG shows that prostaglandin D synthase, an enzyme involved in prostaglandins E2 production is upregulated in female neurons.^{[143,](#page-18-5)[145](#page-18-7)} Prostaglandins E2 is well-known to be involved in pain processing by inducing neuronal sensitization. Indeed, cyclo-oxygenase inhibitors have been widely used in pain management. Importantly, studies in rodent arthritis models have described decreased inflammation in cyclo-oxygenase–deficient mice in females, but not in males.^{[146](#page-18-8)} However, in the same study, pain and sexual dimorphism were not analyzed.

Sex Differences in Microglia and Macrophages

The sensory afferent neuron forms an excitatory synapse with second-order neurons in the dorsal horn of the dorsal spinal cord to initiate transmission in the central nervous system. A sexual dimorphism in pain processing in the spinal cord has been described in the literature, $67,147$ $67,147$ $67,147$ including in animal models of RA (see [Table 1](#page-6-0)). Mogil and colleagues have shown that hypersensitivity in inflammatory and neuropathic pain conditions in mice, can be attributed to distinct immune cell types: microglia in males and T cells in females.^{[148,](#page-18-10)[149](#page-18-11)} Corroborating this finding, the response to intrathecal administration of glial inhibitors are only effec-tive in males and not in females in CAIA mice^{[112](#page-16-19)} (see [Table 1](#page-6-0)) and reinforced a sex difference in pain neuraxial mechanisms. The purinergic receptor P2X4 is known to play key roles in inflammatory response^{[150](#page-18-12)} and is involved in a sex-dependent manner in various pain conditions.^{[151](#page-18-13)} In neuropathic pain, a male-specific upregu-lation of P2X4R has been shown^{[148](#page-18-10)} and an inhibition of spinal P2X4R attenuates pain hypersensitivity in males but not females mice,^{[152](#page-18-14)} which raises the questions of whether microglial purinergic receptors could be involved in pain and sexual dimorphism in RA.

Sex Differences in Pain Processing by the Brain

Other than DRGs or spinal cord changes, supraspinal structures involved with high-order functions, such as anxiety, depression, and cognition, are critical component for pain processing. 63 There is a sex-related difference in brain structures involved in pain perception and modulation that can be explained, at least in part, by sex-hormones. Periaqueductal gray neurons, which project to the rostral ventral medulla and constituting the primary descending pathway of pain inhibition, express sex hormone receptors. AR and $ER\alpha$ immunoreactive neurons were widely distributed in the caudal periaqueductal gray neurons of male rats. Females had significantly fewer AR-positive neurons, although the quantity of $ER\alpha$ was comparable between the sexes.^{[153](#page-18-15)} A preclinical study showed that rostral anterior cingulate cortex, an important brain structure involved in pain affect, is also modulated by estrogen via $ER¹⁵⁴$ $ER¹⁵⁴$ $ER¹⁵⁴$ Considering deep tissue pain in humans, brain imaging studies showed complex brain networks involved in sensory and affective aspects of pain. Under many chronic pain conditions, it was observed greater anterior cingulate cortex activation in females and insular cortex activation in males.[155](#page-19-0) This sexual dimorphism of brain areas activation might contribute to different pain-related responses, which could require different treatment based on sex/gender differences. Although sex hormones have been suggested as key drivers of the sexual dimorphism observed in the expression of pain, an important sociocultural component must be also taking into account to explain the difference of pain experience in humans.

SUMMARY AND FUTURE DIRECTIONS

Epidemiologic and clinical findings demonstrate sex differences in several pain conditions, with women at higher risk of developing chronic pain. The central processing of pain information involving higher order neural functions can explain, at least in part, the sex-related differences in pain experienced by patients with RA. However, there is a dearth in the literature of studies directly addressing sex-associated differences and behavior in rodent models of inflammatory pain. As highlighted by Krock and col-leagues,^{[104](#page-16-0)} nearly 50% of studies focused on pain-like behavior in arthritis were conducted only in male rodents. This observation reinforced that female and especially female/male comparisons in arthritis pain research are still needed. Although RA is an area of intense study, a single rodent model does not fully recapitulate disease. However, current models have been a useful tool to understand the pathophysiology and new targets to treat RA. Therefore, investigations with males and females are needed to advance our understanding of which components of the inflammatory processes that generate pain are subject to sex dimorphisms.

Therapies like nonsteroidal anti-inflammatory drugs have been used to treat pain in RA and in preclinical models of RA. $110,156$ $110,156$ $110,156$ However, is important to note that studies suggest a neuropathic phenotype in RA-associated pain, $110,157$ $110,157$ which would not be adequately treated by NSAIDs. Therefore, examining available treatments used to treat neuropathic pain might be valuable in addressing RA-associated pain. Strikingly, a sexual dimorphism has been seen when it comes to pain management in some an-imal models.^{[158](#page-19-3)} This finding highlights the necessity for understanding the phenotype of arthritis in both males and females to direct an effective treatment. It is noteworthy that, in parallel to biological factors, psychosocial and cultural factors contribute strongly to sexual dimorphism of pain perception in patients, $159-164$ and consequently their response to therapy.

In conclusion, several animal models of RA have contributed our understanding of the pathogenesis of RA and therapeutic management. However, we still need to navigate through RA under a pain perspective, correlating key factors of RA that have not been tested in the pain context and considering both sexes.

CLINICS CARE POINTS

- Inflammatory joint pain exemplified in rheumatoid arthritis is regulated by different processes in males and females.
- Some, but not all, of the sex differences in pain processing are associated with sex hormones and their receptors.
- Sexual dimorphism in pain processing may also lead to sex differences in response to treatment and therapies should be evaluated for differences in efficacy in males and females.

DISCLOSURE

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REFERENCES

- 1. [Tsoucalas G, Sgantzos M. Primary Asthenic gout by Augustin-Jacob Landre-](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref1)[Beauvais in 1800: is this the first description of rheumatoid arthritis? Mediterr](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref1) [J Rheumatol 2017;28\(4\):223–6.](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref1)
- 2. Jokar M, Jokar M. Prevalence of inflammatory rheumatic diseases in a rheumatologic outpatient clinic: analysis of 12626 cases. Rheumatol Res 2018;3(1): 21–7. [https://doi.org/10.22631/rr.2017.69997.1037.](https://doi.org/10.22631/rr.2017.69997.1037)
- 3. WHO scientific group on the burden of musculoskeletal conditions at the start of the new millennium. The burden of musculoskeletal conditions at the start of the new millennium. World Health Organ Tech Rep Ser. 2003;919:i-x, 1-218, back cover. Available at: <http://www.ncbi.nlm.nih.gov/pubmed/14679827>. Accessed January 15, 2018.
- 4. Minichiello E, Semerano L, Boissier MC. Incidence, prévalence et sévérité de la polyarthrite rhumatoïde au XXIe siècle. Rev du Rhum Monogr 2017;84(4): 303–10. [https://doi.org/10.1016/j.monrhu.2017.07.002.](https://doi.org/10.1016/j.monrhu.2017.07.002)
- 5. [Cross M, Smith E, Hoy D, et al. The global burden of rheumatoid arthritis: esti](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref5)[mates from the global burden of disease 2010 study. Ann Rheum Dis 2014;](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref5) [73\(7\):1316–22](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref5).
- 6. [Rudan I, Sidhu S, Papana A, et al. Prevalence of rheumatoid arthritis in low- and](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref6) [middle-income countries: a systematic review and analysis. J Glob Health 2015;](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref6) [5\(1\):010409.](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref6)
- 7. [Smolen JS, Aletaha D, Barton A, et al. Rheumatoid arthritis. Nat Rev Dis Primers](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref7) [2018;4:18001](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref7).
- 8. [Aletaha D, Neogi T, Silman AJ, et al. 2010 Rheumatoid arthritis classification](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref8) [criteria: an American college of rheumatology/European league against rheu](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref8)[matism collaborative initiative. Arthritis Rheum 2010;62\(9\):2569–81.](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref8)
- 9. [van Zanten A, Arends S, Roozendaal C, et al. Presence of anticitrullinated pro](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref9)[tein antibodies in a large population-based cohort from the Netherlands. Ann](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref9) [Rheum Dis 2017;76\(7\):1184–90.](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref9)
- 10. Musset L, Ghillani-Dalbin P. La polyarthrite rhumatoïde: Apport de la biologie au diagnostic et au suivi thérapeutique. Immuno-Analyse Biol Spec 2013;28(5-6): 281–6. [https://doi.org/10.1016/j.immbio.2013.05.001.](https://doi.org/10.1016/j.immbio.2013.05.001)
- 11. Frisell T, Holmqvist M, Källberg H, et al. Familial risks and heritability of rheuma[toid arthritis: role of rheumatoid factor/anti-citrullinated protein antibody status,](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref11) [number and type of affected relatives, sex, and age. Arthritis Rheum 2013;](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref11) [65\(11\):2773–82](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref11).
- 12. [Frisell T, Saevarsdottir S, Askling J. Family history of rheumatoid arthritis: an old](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref12) [concept with new developments. Nat Rev Rheumatol 2016;12\(6\):335–43](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref12).
- 13. [Viatte S, Barton A. Genetics of rheumatoid arthritis susceptibility, severity, and](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref13) [treatment response. Semin Immunopathol 2017;39\(4\):395–408](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref13).
- 14. [Okada Y, Wu D, Trynka G, et al. Genetics of rheumatoid arthritis contributes to](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref14) [biology and drug discovery. Nature 2014;506\(7488\):376–81.](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref14)
- 15. Angelotti F, Parma A, Cafaro G, et al. One year in review 2017: pathogenesis of rheumatoid arthritis. Clin Exp Rheumatol 2017;35(3):0368–78. Available at: <http://www.ncbi.nlm.nih.gov/pubmed/28631608>. Accessed August 1, 2017.
- 16. Richez C, Barnetche T, Schaeverbeke T, Truchetet ME. La polyarthrite rhumatoïde: une physiopathologie mieux connue? Rev du Rhum Monogr 2017; 84(4):311–7. [https://doi.org/10.1016/j.monrhu.2017.07.006.](https://doi.org/10.1016/j.monrhu.2017.07.006)
- 17. [Ai R, Hammaker D, Boyle DL, et al. Joint-specific DNA methylation and tran](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref17)[scriptome signatures in rheumatoid arthritis identify distinct pathogenic pro](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref17)[cesses. Nat Commun 2016;7:11849.](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref17)
- 18. [Hammaker D, Firestein GS. Epigenetics of inflammatory arthritis. Curr Opin](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref18) [Rheumatol 2018;30\(2\):188–96.](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref18)
- 19. [Doody KM, Bottini N, Firestein GS. Epigenetic alterations in rheumatoid arthritis](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref19) [fibroblast-like synoviocytes. Epigenomics 2017;9\(4\):479–92](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref19).
- 20. [Grabiec AM, Korchynskyi O, Tak PP, et al. Histone deacetylase inhibitors sup](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref20)[press rheumatoid arthritis fibroblast-like synoviocyte and macrophage IL-6 pro](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref20)[duction by accelerating mRNA decay. Ann Rheum Dis 2012;71\(3\):424–31](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref20).
- 21. [Mu N, Gu J, Huang T, et al. A novel NF-](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref21)kB/YY1/microRNA-10a regulatory circuit [in fibroblast-like synoviocytes regulates inflammation in rheumatoid arthritis. Sci](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref21) [Rep 2016;6\(1\):20059](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref21).
- 22. [Chang K, Yang SM, Kim SH, et al. Smoking and rheumatoid arthritis. Int J Mol](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref22) [Sci 2014;15\(12\):22279–95](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref22).
- 23. [Costenbader KH, Feskanich D, Mandl LA, et al. Smoking intensity, duration, and](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref23) [cessation, and the risk of rheumatoid arthritis in women. Am J Med 2006;119\(6\):](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref23) [503.e1–9](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref23).
- 24. [Di Giuseppe D, Orsini N, Alfredsson L, et al. Cigarette smoking and smoking](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref24) [cessation in relation to risk of rheumatoid arthritis in women. Arthritis Res Ther](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref24) [2013;15\(2\):R56.](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref24)
- 25. Hedström AK, Stawiarz L, Klareskog L, et al. Smoking and susceptibility to rheumatoid arthritis in a Swedish population-based case–control study. Eur J Epidemiol 2018. [https://doi.org/10.1007/s10654-018-0360-5.](https://doi.org/10.1007/s10654-018-0360-5)
- 26. Källberg H, Ding B, Padyukov L, et al. Smoking is a major preventable risk factor [for rheumatoid arthritis: estimations of risks after various exposures to cigarette](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref26) [smoke. Ann Rheum Dis 2011;70\(3\):508–11](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref26).
- 27. [Lee S-Y, Lee SH, Jhun J, et al. A combination with probiotic complex, zinc, and](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref27) [coenzyme Q10 attenuates autoimmune arthritis by regulation of Th17/Treg bal](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref27)[ance. J Med Food 2018;21\(1\):39–46](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref27).
- 28. [Zhang X, Zhang D, Jia H, et al. The oral and gut microbiomes are perturbed in](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref28) [rheumatoid arthritis and partly normalized after treatment. Nat Med 2015;21\(8\):](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref28) [895–905](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref28).
- 29. [Scher JU, Sczesnak A, Longman RS, et al. Expansion of intestinal Prevotella co](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref29)[pri correlates with enhanced susceptibility to arthritis. Elife 2013;2:e01202](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref29).
- 30. [Khandpur R, Carmona-Rivera C, Vivekanandan-Giri A, et al. NETs are a source](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref30) [of citrullinated autoantigens and stimulate inflammatory responses in rheuma](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref30)[toid arthritis. Sci Transl Med 2013;5\(178\):178ra40](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref30).
- 31. [Sur Chowdhury C, Giaglis S, Walker UA, et al. Enhanced neutrophil extracellular](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref31) [trap generation in rheumatoid arthritis: analysis of underlying signal transduc](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref31)[tion pathways and potential diagnostic utility. Arthritis Res Ther 2014;16\(3\):](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref31) [R122.](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref31)
- 32. [Berthelot J-M, Le Goff B, Neel A, et al. NETose: au carrefour des polyarthrites](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref32) rhumatoïdes, lupus et vascularites. Rev Rhum 2017;84(4):274-81.
- 33. [Demoruelle MK, Harrall KK, Ho L, et al. Anti-citrullinated protein antibodies are](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref33) [associated with neutrophil extracellular traps in the sputum in relatives of rheu](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref33)[matoid arthritis patients. Arthritis Rheumatol 2017;69\(6\):1165–75.](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref33)
- 34. [van](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref34) [Loosdregt](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref34) [J,](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref34) [Rossetti](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref34) [M,](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref34) [Spreafico](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref34) [R,](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref34) [et](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref34) [al.](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref34) [Increased](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref34) [autophagy](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref34) [in](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref34) [CD4](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref34)⁺ [T cells of rheumatoid arthritis patients results in T-cell hyperactivation and](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref34) [apoptosis resistance. Eur J Immunol 2016;46\(12\):2862–70.](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref34)
- 35. [Sparks JA, Iversen MD, Miller Kroouze R, et al. Personalized risk estimator for](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref35) [rheumatoid arthritis \(PRE-RA\) family study: rationale and design for a random](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref35)[ized controlled trial evaluating rheumatoid arthritis risk education to first](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref35)[degree relatives. Contemp Clin Trials 2014;39\(1\):145–57.](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref35)
- 36. [Ferucci ED, Templin DW, Lanier AP. Rheumatoid arthritis in American Indians](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref36) [and Alaska natives: a review of the literature. Semin Arthritis Rheum 2005;](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref36) [34\(4\):662–7](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref36).
- 37. [McMichael AJ, Sasazuki T, McDevitt HO, et al. Increased frequency of HLA-Cw3](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref37) [and HLA-Dw4 in rheumatoid arthritis. Arthritis Rheum 1977;20\(5\):1037–42.](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref37)
- 38. [Stastny P. Association of the B-Cell Alloantigen DRw4 with rheumatoid arthritis.](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref38) [N Engl J Med 1978;298\(16\):869–71](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref38).
- 39. Gregersen PK, Silver J, Winchester RJ. The shared epitope hypothesis. An approach to understanding the molecular genetics of susceptibility to rheumatoid arthritis. Arthritis Rheum 1987;30(11):1205–13. Available at: [http://www.](http://www.ncbi.nlm.nih.gov/pubmed/2446635) [ncbi.nlm.nih.gov/pubmed/2446635](http://www.ncbi.nlm.nih.gov/pubmed/2446635). Accessed March 5, 2018.
- 40. [Raychaudhuri S, Sandor C, Stahl EA, et al. Five amino acids in three HLA pro](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref40)[teins explain most of the association between MHC and seropositive rheumatoid](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref40) [arthritis. Nat Genet 2012;44\(3\):291–6.](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref40)
- 41. [Ling SF, Viatte S, Lunt M, et al. HLA-DRB1 amino acid positions 11/13, 71, and](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref41) [74 are associated with inflammation level, disease activity, and the health](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref41) [assessment questionnaire score in patients with inflammatory polyarthritis.](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref41) [Arthritis Rheumatol 2016;68\(11\):2618–28.](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref41)
- 42. Piñero J, Queralt-Rosinach N, Bravo À[, et al. DisGeNET: a discovery platform for](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref42) [the dynamical exploration of human diseases and their genes. Database \(Ox](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref42)[ford\) 2015;2015:bav028](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref42).
- 43. Stolt P, Källberg H, Lundberg I, et al. Silica exposure is associated with [increased risk of developing rheumatoid arthritis: results from the Swedish](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref43) [EIRA study. Ann Rheum Dis 2005;64\(4\):582–6](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref43).
- 44. [Deane KD, Demoruelle MK, Kelmenson LB, et al. Genetic and environmental](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref44) [risk factors for rheumatoid arthritis. Best Pract Res Clin Rheumatol 2017;](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref44) [31\(1\):3–18](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref44).
- 45. Vaahtovuo J, MUNUKKA E, KORKEAMÄKI M, et al. Fecal Microbiota in Early Rheumatoid Arthritis. J Rheumatol 2008;35(8). Available at: [http://www.jrheum.](http://www.jrheum.org/content/35/8/1500.long) [org/content/35/8/1500.long.](http://www.jrheum.org/content/35/8/1500.long) Accessed January 19, 2018.
- 46. Gul'neva MI, Noskov SM. Colonic microbial biocenosis in rheumatoid arthritis. Klin Med (Mosk) 2011;89(4):45–8. Available at: [http://www.ncbi.nlm.nih.gov/](http://www.ncbi.nlm.nih.gov/pubmed/21932563) [pubmed/21932563.](http://www.ncbi.nlm.nih.gov/pubmed/21932563) Accessed January 19, 2018.
- 47. [Liu X, Zou Q, Zeng B, et al. Analysis of fecal lactobacillus community structure in](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref47) [patients with early rheumatoid arthritis. Curr Microbiol 2013;67\(2\):170–6.](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref47)
- 48. Khalifa O, Pers YM, Ferreira R, et al. X-linked miRNAs associated with gender differences in rheumatoid arthritis. Int J Mol Sci 2016;17(11). [https://doi.org/](https://doi.org/10.3390/ijms17111852) [10.3390/ijms17111852.](https://doi.org/10.3390/ijms17111852)
- 49. [Ortona E, Pierdominici M, Maseli A, et al. Sex-based differences in autoimmune](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref49) [diseases. Ann Ist Super Sanita 2016;52\(2\):205–12.](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref49)
- 50. [Jawaheer D, Lum RF, Gregersen PK, et al. Influence of male sex on disease](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref50) [phenotype in familial rheumatoid arthritis. Arthritis Rheum 2006;54\(10\):3087–94.](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref50)
- 51. [Hughes GC, Choubey D. Modulation of autoimmune rheumatic diseases by](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref51) [oestrogen and progesterone. Nat Rev Rheumatol 2014;10\(12\):740–51.](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref51)
- 52. [Chen H, Wang J, Zhou W, et al. Breastfeeding and risk of rheumatoid arthritis: a](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref52) [systematic review and metaanalysis. J Rheumatol 2015;42\(9\):1563–9.](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref52)
- 53. Alpízar-Rodríguez D, Pluchino N, Canny G, Gabay C, Finckh A. The role of female hormonal factors in the development of rheumatoid arthritis. Rheumatol (United Kingdom) 2017;56(8):1254–63. [https://doi.org/10.1093/rheumatology/](https://doi.org/10.1093/rheumatology/kew318) [kew318](https://doi.org/10.1093/rheumatology/kew318).
- 54. Dimitrijević M, Arsenović-Ranin N, Kosec D, et al. Sex differences in Tfh cell help to B cells contribute to sexual dimorphism in severity of rat collagen-induced arthritis. Sci Rep 2020;10(1). [https://doi.org/10.1038/s41598-020-58127-y.](https://doi.org/10.1038/s41598-020-58127-y)
- 55. McWilliams DF, Walsh DA. Pain mechanisms in rheumatoid arthritis. Suppl 1(5). Clin Exp Rheumatol 2017;35:94–101. Available at: [http://www.ncbi.nlm.nih.gov/](http://www.ncbi.nlm.nih.gov/pubmed/28967354) [pubmed/28967354](http://www.ncbi.nlm.nih.gov/pubmed/28967354). Accessed July 3, 2018.
- 56. [van Delft MAM, Huizinga TWJ. An overview of autoantibodies in rheumatoid](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref56) [arthritis. J Autoimmun 2020;110:102392](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref56).
- 57. [Lee YC, Cui J, Lu B, et al. Pain persists in DAS28 rheumatoid arthritis remission](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref57) [but not in ACR/EULAR remission: a longitudinal observational study. Arthritis](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref57) [Res Ther 2011;13\(3\):R83](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref57).
- 58. [Boeters DM, Burgers LE, Toes REM, et al. Does immunological remission,](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref58) [defined as disappearance of autoantibodies, occur with current treatment stra](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref58)[tegies? A long-term follow-up study in rheumatoid arthritis patients who](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref58) [achieved sustained DMARD-free status. Ann Rheum Dis 2019;78\(11\):1497–504.](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref58)
- 59. [Walsh DA, McWilliams DF. Mechanisms, impact and management of pain in](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref59) [rheumatoid arthritis. Nat Rev Rheumatol 2014;10\(10\):581–92](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref59).
- 60. Catrina AI, Svensson CI, Malmström V, et al. Mechanisms leading from systemic [autoimmunity to joint-specific disease in rheumatoid arthritis. Nat Rev Rheuma](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref60)[tol 2017;13\(2\):79–86.](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref60)
- 61. [Bas DB, Su J, Wigerblad G, et al. Pain in rheumatoid arthritis: models and mech](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref61)[anisms. Pain Manag 2016;6\(3\):265–84](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref61).
- 62. Sarzi-Puttini P, Salaffi F, Di Franco M, et al. Pain in rheumatoid arthritis: a critical review. Reumatismo 2014;66(1):18–27. Available at: [http://www.ncbi.nlm.nih.](http://www.ncbi.nlm.nih.gov/pubmed/24938192) [gov/pubmed/24938192.](http://www.ncbi.nlm.nih.gov/pubmed/24938192) Accessed April 27, 2018.
- 63. Gonçalves dos Santos G, Delay L, Yaksh TL, et al. Neuraxial cytokines in pain [states. Front Immunol 2020;10:3061.](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref63)
- 64. [Yaksh TL, Hunt MA, dos Santos GG. Development of new analgesics: an answer](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref64) [to opioid epidemic. Trends Pharmacol Sci 2018;39\(12\):1000–2](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref64).
- 65. [Torrance N, Smith BH, Bennett MI, et al. The epidemiology of chronic pain of](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref65) [predominantly neuropathic origin. results from a general population survey.](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref65) [J Pain 2006;7\(4\):281–9](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref65).
- 66. [Fillingim RB, King CD, Ribeiro-Dasilva MC, et al. Sex, gender, and pain: a review](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref66) [of recent clinical and experimental findings. J Pain 2009;10\(5\):447–85.](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref66)
- 67. [Mapplebeck JCS, Beggs S, Salter MW. Sex differences in pain: a tale of two im](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref67)[mune cells. Pain 2016;157:S2–6](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref67).
- 68. [Aloisi AM. Why we still need to speak about sex differences and sex hormones](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref68) [in pain. Pain Ther 2017;6\(2\):111–4.](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref68)
- 69. [Radawski C, Genovese MC, Hauber B, et al. Patient perceptions of unmet med](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref69)[ical need in rheumatoid arthritis: a cross-sectional survey in the USA. Rheumatol](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref69) [Ther 2019;6\(3\):461–71](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref69).
- 70. [Taylor P, Manger B, Alvaro-Gracia J, et al. Patient perceptions concerning pain](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref70) [management in the treatment of rheumatoid arthritis. J Int Med Res 2010;38\(4\):](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref70) [1213–24.](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref70)
- 71. [Heiberg T, Kvien TK. Preferences for improved health examined in 1,024 pa](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref71)[tients with rheumatoid arthritis: pain has highest priority. Arthritis Rheum 2002;](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref71) [47\(4\):391–7](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref71).
- 72. Bergstra SA, Allaart CF. What is the optimal target for treat-to-target strategies in rheumatoid arthritis? Curr Opin Rheumatol 2018;1. [https://doi.org/10.1097/BOR.](https://doi.org/10.1097/BOR.0000000000000484) [0000000000000484.](https://doi.org/10.1097/BOR.0000000000000484)
- 73. [van Baarsen LGM, de Hair MJH, Ramwadhdoebe TH, et al. The cellular compo](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref73)[sition of lymph nodes in the earliest phase of inflammatory arthritis. Ann Rheum](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref73) [Dis 2013;72\(8\):1420–4](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref73).
- 74. [de Hair MJH, van de Sande MGH, Ramwadhdoebe TH, et al. Features of the](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref74) [synovium of individuals at risk of developing rheumatoid arthritis: implications](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref74) [for understanding preclinical rheumatoid arthritis. Arthritis Rheumatol 2014;](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref74) [66\(3\):513–22](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref74).
- 75. [Das S, Padhan P. An overview of the extraarticular involvement in rheumatoid](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref75) [arthritis and its management. J Pharmacol Pharmacother 2017;8\(3\):81–6](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref75).
- 76. [Deane KD, Norris JM, Holers VM. Preclinical rheumatoid arthritis: identification,](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref76) [evaluation, and future directions for investigation. Rheum Dis Clin North Am](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref76) [2010;36\(2\):213–41.](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref76)
- 77. Kurki P, Aho K, Palosuo T, et al. Immunopathology of rheumatoid arthritis. Antikeratin antibodies precede the clinical disease. Arthritis Rheum 1992;35(8): 914–7. Available at: [http://www.ncbi.nlm.nih.gov/pubmed/1379430.](http://www.ncbi.nlm.nih.gov/pubmed/1379430) Accessed April 19, 2018.
- 78. [Nielen MMJ, van Schaardenburg D, Reesink HW, et al. Specific autoantibodies](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref78) [precede the symptoms of rheumatoid arthritis: a study of serial measurements in](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref78) [blood donors. Arthritis Rheum 2004;50\(2\):380–6.](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref78)
- 79. [van de Stadt LA, de Koning MHMT, van de Stadt RJ, et al. Development of the](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref79) [anti-citrullinated protein antibody repertoire prior to the onset of rheumatoid](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref79) [arthritis. Arthritis Rheum 2011;63\(11\):3226–33](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref79).
- 80. [Shi J, Knevel R, Suwannalai P, et al. Autoantibodies recognizing carbamylated](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref80) [proteins are present in sera of patients with rheumatoid arthritis and predict joint](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref80) [damage. Proc Natl Acad Sci U S A 2011;108\(42\):17372–7](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref80).
- 81. [Shi J, van Steenbergen HW, van Nies JAB, et al. The specificity of anti](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref81)[carbamylated protein antibodies for rheumatoid arthritis in a setting of early](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref81) [arthritis. Arthritis Res Ther 2015;17:339.](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref81)
- 82. [Shi J, Van De Stadt LA, Levarht EWN, et al. Anti-carbamylated protein \(anti-](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref82)[CarP\) antibodies precede the onset of rheumatoid arthritis. Ann Rheum Dis](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref82) [2014;73\(4\):780–3.](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref82)
- 83. [Thiele GM, Duryee MJ, Anderson DR, et al. Malondialdehyde-acetaldehyde ad](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref83)[ducts and anti-malondialdehyde-acetaldehyde antibodies in rheumatoid](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref83) [arthritis. Arthritis Rheumatol 2015;67\(3\):645–55.](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref83)
- 84. [Mikuls TR, Duryee MJ, Rahman R, et al. Enrichment of malondialdehyde–](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref84) [acetaldehyde antibody in the rheumatoid arthritis joint. Rheumatology 2017;](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref84) [56\(10\):1794–803](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref84).
- 85. [Wigerblad G, Bas DB, Fernades-Cerqueira C, et al. Autoantibodies to citrulli](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref85)[nated proteins induce joint pain independent of inflammation via a](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref85) [chemokine-dependent mechanism. Ann Rheum Dis 2016;75\(4\):730–8](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref85).
- 86. [Harre U, Georgess D, Bang H, et al. Induction of osteoclastogenesis and bone](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref86) [loss by human autoantibodies against citrullinated vimentin. J Clin Invest 2012;](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref86) [122\(5\):1791–802.](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref86)
- 87. [Scherer HU, van der Woude D, Ioan-Facsinay A, et al. Glycan profiling of anti](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref87)[citrullinated protein antibodies isolated from human serum and synovial fluid.](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref87) [Arthritis Rheum 2010;62\(6\):1620–9.](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref87)
- 88. [Rombouts Y, Ewing E, van de Stadt LA, et al. Anti-citrullinated protein antibodies](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref88) [acquire a pro-inflammatory Fc glycosylation phenotype prior to the onset of](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref88) [rheumatoid arthritis. Ann Rheum Dis 2015;74\(1\):234–41](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref88).
- 89. [Pfeifle R, Rothe T, Ipseiz N, et al. Regulation of autoantibody activity by the IL-](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref89)[23-TH17 axis determines the onset of autoimmune disease. Nat Immunol 2017;](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref89) [18\(1\):104–13.](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref89)
- 90. [Molendijk M, Hazes JM, Lubberts E. From patients with arthralgia, pre-RA and](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref90) [recently diagnosed RA: what is the current status of understanding RA patho](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref90)[genesis? RMD Open 2018;4\(1\):e000256.](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref90)
- 91. [Deane KD, O'Donnell CI, Hueber W, et al. The number of elevated cytokines and](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref91) [chemokines in preclinical seropositive rheumatoid arthritis predicts time to diag](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref91)[nosis in an age-dependent manner. Arthritis Rheum 2010;62\(11\):3161–72.](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref91)
- 92. Kokkonen H, Söderström I, Rocklöv J, et al. Up-regulation of cytokines and che[mokines predates the onset of rheumatoid arthritis. Arthritis Rheum 2010;62\(2\):](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref92) [383–91.](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref92)
- 93. [Chalan P, Bijzet J, van den Berg A, et al. Analysis of serum immune markers in](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref93) [seropositive and seronegative rheumatoid arthritis and in high-risk seropositive](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref93) [arthralgia patients. Sci Rep 2016;6:26021.](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref93)
- 94. [Lee YC, Lu B, Boire G, et al. Incidence and predictors of secondary fibromyalgia](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref94) [in an early arthritis cohort. Ann Rheum Dis 2013;72\(6\):949–54.](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref94)
- 95. [Duffield SJ, Miller N, Zhao S, et al. Concomitant fibromyalgia complicating](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref95) [chronic inflammatory arthritis: a systematic review and meta-analysis. Rheuma](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref95)[tology \(Oxford\) 2018;57\(8\):1453–60.](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref95)
- 96. Bodnar RJ, Romero MT, Kramer E. Organismic variables and pain inhibition: roles of gender and aging. Brain Res Bull 1988. [https://doi.org/10.1016/0361-](https://doi.org/10.1016/0361-9230(88)90032-9) [9230\(88\)90032-9.](https://doi.org/10.1016/0361-9230(88)90032-9)
- 97. [Romero MT, Kepler KL, Bodnar RJ. Gender determinants of opioid mediation of](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref97) [swim analgesia in rats. Pharmacol Biochem Behav 1988;29\(4\):705–9.](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref97)
- 98. Fillingim RB, Maixner W. Gender differences in the responses to noxious stimuli. Pain Forum 1995;4(4):209–21. [https://doi.org/10.1016/S1082-3174\(11\)80022-X.](https://doi.org/10.1016/S1082-3174(11)80022-X)
- 99. [Melchior M, Poisbeau P, Gaumond I, et al. Insights into the mechanisms and the](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref99) [emergence of sex-differences in pain. Neuroscience 2016;338:63–80.](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref99)
- 100. [Mogil JS. Qualitative sex differences in pain processing: emerging evidence of](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref100) [a biased literature. Nat Rev Neurosci 2020;21\(7\):353–65](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref100).
- 101. Rovner GS, Sunnerhagen KS, Björkdahl A, et al. Chronic pain and sex[differences; women accept and move, while men feel blue. PLoS One 2017;](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref101) [12\(4\):1–12.](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref101)
- 102. Fischer BD, Adeyemo A, O'Leary ME, et al. Animal models of rheumatoid pain: experimental systems and insights. Arthritis Res Ther 2017;19(1). [https://doi.](https://doi.org/10.1186/s13075-017-1361-6) [org/10.1186/s13075-017-1361-6.](https://doi.org/10.1186/s13075-017-1361-6)
- 103. [Choudhary N, Bhatt LK, Prabhavalkar KS. Experimental animal models for rheu](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref103)[matoid arthritis. Immunopharmacol Immunotoxicol 2018;40\(3\):193–200.](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref103)
- 104. [Krock E, Jurczak A, Svensson CI. Pain pathogenesis in rheumatoid arthritis](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref104)[what have we learned from animal models? Pain 2018;159\(Suppl 1\):S98–109.](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref104)
- 105. [Cook CD, Nickerson MD. Nociceptive sensitivity and opioid antinociception and](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref105) [antihyperalgesia in Freund's adjuvant-induced arthritic male and female rats.](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref105) [J Pharmacol Exp Ther 2005;313\(1\):449–59](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref105).
- 106. Chillingworth NL, Donaldson LF. Characterisation of a Freund's complete adjuvant-induced model of chronic arthritis in mice. J Neurosci Methods 2003;128(1–2):45–52. Available at: [http://www.ncbi.nlm.nih.gov/pubmed/](http://www.ncbi.nlm.nih.gov/pubmed/12948547) [12948547](http://www.ncbi.nlm.nih.gov/pubmed/12948547). Accessed September 21, 2018.
- 107. [Gauldie SD, McQueen DS, Clarke CJ, et al. A robust model of adjuvant-induced](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref107) [chronic unilateral arthritis in two mouse strains. J Neurosci Methods 2004;](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref107) [139\(2\):281–91](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref107).
- 108. [Inglis JJ, Notley CA, Essex D, et al. Collagen-induced arthritis as a model of hy](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref108)[peralgesia: functional and cellular analysis of the analgesic actions of tumor ne](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref108)[crosis factor blockade. Arthritis Rheum 2007;56\(12\):4015–23.](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref108)
- 109. [Kouskoff V, Korganow AS, Duchatelle V, et al. Organ-specific disease provoked](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref109) [by systemic autoimmunity. Cell 1996;87\(5\):811–22](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref109).
- 110. [Christianson CA, Corr M, Firestein GS, et al. Characterization of the acute and](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref110) [persistent pain state present in K/BxN serum transfer arthritis. Pain 2010;](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref110) [151\(2\):394–403](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref110).
- 111. [Ashraf S, Bouhana KS, Pheneger J, et al. Selective inhibition of tropomyosin](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref111)[receptor-kinase A \(TrkA\) reduces pain and joint damage in two rat models of in](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref111)[flammatory arthritis. Arthritis Res Ther 2016;18\(1\):97](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref111).
- 112. [Fernandez-Zafra T, Gao T, Jurczak A, et al. Exploring the transcriptome of resi](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref112)[dent spinal microglia after collagen antibody-induced arthritis. Pain 2019;](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref112) [160\(1\):224–36](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref112).
- 113. [Woller SA, Ocheltree C, Wong SY, et al. Neuraxial TNF and IFN-beta co-modu](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref113)[late persistent allodynia in arthritic mice. Brain Behav Immun 2019;76:151–8](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref113).
- 114. Sun R-Q, Lawand NB, Willis WD. The role of calcitonin gene-related peptide (CGRP) in the generation and maintenance of mechanical allodynia and hyperalgesia in rats after intradermal injection of capsaicin. Pain 2003;104(1–2): 201–8. Available at: [http://www.ncbi.nlm.nih.gov/pubmed/12855330.](http://www.ncbi.nlm.nih.gov/pubmed/12855330) Accessed August 17, 2018.
- 115. Vincent K, Tracey I. Hormones and their interaction with the pain experience. Rev Pain 2008. <https://doi.org/10.1177/204946370800200206>.
- 116. [Tang MW, Garcia S, Gerlag DM, et al. Insight into the endocrine system and the](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref116) [immune system: a review of the inflammatory role of prolactin in rheumatoid](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref116) [arthritis and psoriatic arthritis. Front Immunol 2017;8\(JUN\):23](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref116).
- 117. [Liu L, Jia J, Jiang M, et al. High susceptibility to collagen-induced arthritis in](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref117) [mice with progesterone receptors selectively inhibited in osteoprogenitor cells.](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref117) [Arthritis Res Ther 2020;22\(1\):1–12](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref117).
- 118. Bupp MRG, Jorgensen TN. Androgen-induced immunosuppression. Front Immunol 2018;9(APR):1. [https://doi.org/10.3389/fimmu.2018.00794.](https://doi.org/10.3389/fimmu.2018.00794)
- 119. [Yang DD, Krasnova A, Nead KT, et al. Androgen deprivation therapy and risk of](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref119) [rheumatoid arthritis in patients with localized prostate cancer. Ann Oncol 2018;](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref119) [29\(2\):386–91](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref119).
- 120. [Baillargeon J, Al Snih S, Raji MA, et al. Hypogonadism and the risk of rheumatic](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref120) [autoimmune disease. Clin Rheumatol 2016;35\(12\):2983–7](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref120).
- 121. Keast JR, Gleeson RJ. Androgen receptor immunoreactivity is present in primary sensory neurons of male rats. Neuroreport 1998. [https://doi.org/10.1097/](https://doi.org/10.1097/00001756-199812210-00025) [00001756-199812210-00025.](https://doi.org/10.1097/00001756-199812210-00025)
- 122. Fanton LE, Macedo CG, Torres-Chávez KE, et al. Activational action of testosterone on androgen receptors protects males preventing temporomandibular joint pain. Pharmacol Biochem Behav 2017. [https://doi.org/10.1016/j.pbb.](https://doi.org/10.1016/j.pbb.2016.07.005) [2016.07.005.](https://doi.org/10.1016/j.pbb.2016.07.005)
- 123. Fischer L, Clemente JT, Tambeli CH. The protective role of testosterone in the development of temporomandibular joint pain. J Pain 2007. [https://doi.org/10.](https://doi.org/10.1016/j.jpain.2006.12.007) [1016/j.jpain.2006.12.007.](https://doi.org/10.1016/j.jpain.2006.12.007)
- 124. Lesnak JB, Inoue S, Lima L, et al. Testosterone protects against the development of widespread muscle pain in mice. Pain 2020. [https://doi.org/10.1097/j.](https://doi.org/10.1097/j.pain.0000000000001985) [pain.0000000000001985](https://doi.org/10.1097/j.pain.0000000000001985).
- 125. Kato Y, Shigehara K, Kawaguchi S, et al. Efficacy of testosterone replacement therapy on pain in hypogonadal men with chronic pain syndrome: a subanalysis of a prospective randomised controlled study in Japan (EARTH study). Andrologia 2020. [https://doi.org/10.1111/and.13768.](https://doi.org/10.1111/and.13768)
- 126. [Gold MS, Flake NM. Inflammation-mediated hyperexcitability of sensory neu](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref126)[rons. Neurosignals 2005;14\(4\):147–57](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref126).
- 127. Cho T, Chaban VV. Expression of P2X3 and TRPV1 receptors in primary sensory neurons from estrogen receptors- α and estrogen receptor- β knockout mice. Neuroreport 2012. [https://doi.org/10.1097/WNR.0b013e328353fabc.](https://doi.org/10.1097/WNR.0b013e328353fabc)
- 128. Taleghany N, Sarajari S, DonCarlos LL, et al. Differential expression of estrogen receptor alpha and beta in rat dorsal root ganglion neurons. J Neurosci Res 1999. [https://doi.org/10.1002/\(SICI\)1097-4547\(19990901\)57:5<603::AID-](https://doi.org/10.1002/(SICI)1097-4547(19990901)57:5<603::AID-JNR3>3.0.CO;2-R)[JNR3>3.0.CO;2-R.](https://doi.org/10.1002/(SICI)1097-4547(19990901)57:5<603::AID-JNR3>3.0.CO;2-R)
- 129. Lu Y, Jiang Q, Yu L, et al. 17 β -estradiol rapidly attenuates P2X3 receptormediated peripheral pain signal transduction via ERa and GPR30. Endocrinology 2013. [https://doi.org/10.1210/en.2012-2119.](https://doi.org/10.1210/en.2012-2119)
- 130. Ma B, hua Yu L, Fan J, et al. Estrogen modulation of peripheral pain signal transduction: involvement of P2X3 receptors. Purinergic Signal 2011. [https://doi.org/](https://doi.org/10.1007/s11302-010-9212-9) [10.1007/s11302-010-9212-9.](https://doi.org/10.1007/s11302-010-9212-9)
- 131. Chaban V, Li J, McDonald JS, et al. Estradiol attenuates the adenosine triphosphate-induced increase of intracellular calcium through group II metabotropic glutamate receptors in rat dorsal root ganglion neurons. J Neurosci Res 2011. [https://doi.org/10.1002/jnr.22718.](https://doi.org/10.1002/jnr.22718)
- 132. Cho T, Chaban VV. Interaction Between P2X3 and Oestrogen Receptor (ER)a/ ER_B in ATP-Mediated Calcium Signalling In Mice Sensory Neurones. J Neuroendocrinol 2012;24(5):789–97. [https://doi.org/10.1111/j.1365-2826.2011.02272.x.](https://doi.org/10.1111/j.1365-2826.2011.02272.x)
- 133. Xu S, Cheng Y, Keast JR, et al. 17 β -estradiol activates estrogen receptor β -signalling and inhibits transient receptor potential vanilloid receptor 1 activation by capsaicin in adult rat nociceptor neurons. Endocrinology 2008. [https://doi.org/](https://doi.org/10.1210/en.2008-0278) [10.1210/en.2008-0278](https://doi.org/10.1210/en.2008-0278).
- 134. Nazarian A, Tenayuca JM, Almasarweh F, et al. Sex differences in formalinevoked primary afferent release of substance P. Eur J Pain 2014. [https://doi.](https://doi.org/10.1002/j.1532-2149.2013.00346.x) [org/10.1002/j.1532-2149.2013.00346.x.](https://doi.org/10.1002/j.1532-2149.2013.00346.x)
- 135. Sohrabji F, Miranda RC, Toran-Allerand CD. Estrogen differentially regulates estrogen and nerve growth factor receptor mRNAs in adult sensory neurons. J Neurosci 1994. <https://doi.org/10.1523/jneurosci.14-02-00459.1994>.
- 136. Liuzzi FJ, Scoville SA, Bufton SM. Long-term estrogen replacement coordinately decreases trkA and β -PPT mRNA levels in dorsal root ganglion neurons. Exp Neurol 1999. <https://doi.org/10.1006/exnr.1998.6999>.
- 137. Lanlua P, Decorti F, Gangula PRR, et al. Female steroid hormones modulate receptors for nerve growth factor in rat dorsal root ganglia. Biol Reprod 2001. [https://doi.org/10.1095/biolreprod64.1.331.](https://doi.org/10.1095/biolreprod64.1.331)
- 138. Garrett NE, Mapp PI, Cruwys SC, et al. Role of substance P in inflammatory arthritis. Ann Rheum Dis 1992. [https://doi.org/10.1136/ard.51.8.1014.](https://doi.org/10.1136/ard.51.8.1014)
- 139. Keeble JE, Brain SD. A role for substance P in arthritis? Neurosci Lett 2004. [https://doi.org/10.1016/j.neulet.2003.12.020.](https://doi.org/10.1016/j.neulet.2003.12.020)
- 140. Denk F, Bennett DL, McMahon SB. Nerve growth factor and pain mechanisms. Annu Rev Neurosci April 2017. [https://doi.org/10.1146/annurev-neuro-072116-](https://doi.org/10.1146/annurev-neuro-072116-031121) [031121](https://doi.org/10.1146/annurev-neuro-072116-031121).
- 141. [Seidel MF, Herguijuela M, Forkert R, et al. Nerve growth factor in rheumatic dis](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref141)[eases. Semin Arthritis Rheum 2010;40\(2\):109–26](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref141).
- 142. Yang Y, Ozawa H, Lu H, et al. Immunocytochemical analysis of sex differences in calcitonin gene- related peptide in the rat dorsal root ganglion, with special reference to estrogen and its receptor. Brain Res 1998. [https://doi.org/10.](https://doi.org/10.1016/S0006-8993(98)00021-3) [1016/S0006-8993\(98\)00021-3.](https://doi.org/10.1016/S0006-8993(98)00021-3)
- 143. [Patil M, Belugin S, Mecklenburg J, et al. Prolactin regulates pain responses via a](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref143) [female-selective nociceptor-specific mechanism. iScience 2019;20:449–65.](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref143)
- 144. [Tang MW, Reedquist KA, Garcia S, et al. The prolactin receptor is expressed in](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref144) [rheumatoid arthritis and psoriatic arthritis synovial tissue and contributes to](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref144) [macrophage activation. Rheumatology \(Oxford\) 2016;55\(12\):2248–59](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref144).
- 145. Tavares-Ferreira D, Ray PR, Sankaranarayanan I, et al. Sex differences in nociceptor translatomes contribute to divergent prostaglandin signaling in male and female mice Short title: sex differences in mouse nociceptor translatomes. bio-Rxiv 2020. [https://doi.org/10.1101/2020.07.31.231753.](https://doi.org/10.1101/2020.07.31.231753)
- 146. [Chillingworth NL, Morham SG, Donaldson LF. Sex differences in inflammation](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref146) [and inflammatory pain in cyclooxygenase-deficient mice. Am J Physiol Regul In](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref146)[tegr Comp Physiol 2006;291\(2\):R327–34](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref146).
- 147. Dimitrijević M, Arsenović-Ranin N, Kosec D, et al. Sex differences in Tfh cell help [to B cells contribute to sexual dimorphism in severity of rat collagen-induced](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref147) [arthritis. Sci Rep 2020;10\(1\):1–15](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref147).
- 148. [Sorge RE, Mapplebeck JCS, Rosen S, et al. Different immune cells mediate me](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref148)[chanical pain hypersensitivity in male and female mice. Nat Neurosci 2015;](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref148) [18\(8\):1081–3](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref148).
- 149. [Sorge RE, LaCroix-Fralish ML, Tuttle AH, et al. Spinal cord toll-like receptor 4](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref149) [mediates inflammatory and neuropathic hypersensitivity in male but not female](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref149) [mice. J Neurosci 2011;31\(43\):15450–4](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref149).
- 150. Zhang W j, Luo H l, Zhu Z m. The role of P2X4 receptors in chronic pain: a potential pharmacological target. Biomed Pharmacother 2020;129. [https://doi.org/](https://doi.org/10.1016/j.biopha.2020.110447) [10.1016/j.biopha.2020.110447](https://doi.org/10.1016/j.biopha.2020.110447).
- 151. [Halievski K, Ghazisaeidi S, Salter MW. Sex-dependent mechanisms of chronic](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref151) [pain: a focus on microglia and P2X4R. J Pharmacol Exp Ther 2020;120:265017](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref151).
- 152. [Mapplebeck JCS, Dalgarno R, Tu YS, et al. Microglial P2X4R-evoked pain hy](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref152)[persensitivity is sexually dimorphic in rats. Pain 2018;159\(9\):1752–63](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref152).
- 153. Loyd DR, Murphy AZ. Androgen and estrogen (α) receptor localization on periaqueductal gray neurons projecting to the rostral ventromedial medulla in the male and female rat. J Chem Neuroanat 2008. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.jchemneu.2008.08.001) [jchemneu.2008.08.001.](https://doi.org/10.1016/j.jchemneu.2008.08.001)
- 154. Xiao X, Yang Y, Zhang Y, et al. Estrogen in the anterior cingulate cortex contributes to pain-related aversion. Cereb Cortex 2013. [https://doi.org/10.1093/](https://doi.org/10.1093/cercor/bhs201) [cercor/bhs201.](https://doi.org/10.1093/cercor/bhs201)
- 155. [Traub RJ, Ji Y. Sex differences and hormonal modulation of deep tissue pain.](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref155) [Front Neuroendocrinol 2013;34\(4\):350–66.](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref155)
- 156. Crofford LJ. Use of NSAIDs in treating patients with arthritis. Arthritis Res Ther 2013;15:S2. [https://doi.org/10.1186/ar4174.](https://doi.org/10.1186/ar4174) Suppl 3(Suppl 3).
- 157. [Christianson CA, Corr M, Yaksh TL, et al. K/BxN serum transfer arthritis as a](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref157) [model of inflammatory joint pain. Methods Mol Biol 2012;851:249–60.](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref157)
- 158. [Hurley RW, Adams MCB. Sex, gender, and pain: an overview of a complex field.](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref158) [Anesth Analg 2008;107\(1\):309–17](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref158).
- 159. [Bartley EJ, Palit S. Gender and pain. Curr Anesthesiol Rep 2016;6\(4\):344–53](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref159).
- 160. [Bartley EJ, Fillingim RB. Sex differences in pain: a brief review of clinical and](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref160) [experimental findings. Br J Anaesth 2013;111\(1\):52–8](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref160).
- 161. [Berkley KJ, Zalcman SS, Simon VR. Sex and gender differences in pain and](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref161) [inflammation: a rapidly maturing field. Am J Physiol Regul Integr Comp Physiol](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref161) [2006;291\(2\):241–4](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref161).
- 162. Greenspan JD, Craft RM, LeResche L, et al. Studying sex and gender differences in pain and analgesia: A consensus report. Pain 2007;132(SUPPL. 1): S26. <https://doi.org/10.1016/j.pain.2007.10.014>.
- 163. [Kuba T, Quinones-Jenab V. The role of female gonadal hormones in behavioral](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref163) [sex differences in persistent and chronic pain: clinical versus preclinical studies.](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref163) [Brain Res Bull 2005;66\(3\):179–88](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref163).
- 164. [Strassman AM, Mineta Y, Vos BP. Distribution of fos-like immunoreactivity in the](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref164) [medullary and upper cervical dorsal horn produced by stimulation of dural](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref164) [blood vessels in the rat. J Neurosci 1994;14\(6\):3725–35](http://refhub.elsevier.com/S0889-857X(20)30141-1/sref164).