



High expression of HOXA5 is associated with poor prognosis in acute myeloid leukemia

You Yang^{a,b}, Fangfang Zhong^{a,b}, Xiaoming Huang^a, Na Zhang^a, Jingjing Du^a, Ze Long^a, Bowen Zheng^a, Wanjun Lin^a, Wenjun Liu^{b,*}, Wenzhe Ma^{a,**}

^a State Key Laboratory of Quality Research in Chinese Medicine, Macau University of Science and Technology, Macau, China

^b Department of Pediatrics, Affiliated Hospital of Southwest Medical University, Sichuan Clinical Research Center for Birth Defects, Luzhou, Sichuan, China

A B S T R A C T

Background: HOXA5 is considered as an oncogene in many tumors. This study investigated the HOXA5 expression in Chinese acute myeloid leukemia (AML) patients and evaluated the predictive significance of HOXA5 with a single-center retrospective study. **Methods:** We investigated the expression pattern and prognostic value of HOXA5 in patients with AML through by using a series of databases and various datasets, including the ONCOMINE, TCGA, and STRING datasets. The bone marrow samples of 53 newly diagnosed AML patients (non-M3 subtype) and 19 benign individuals were collected in our center. HOXA5 mRNA expression levels were detected by real-time qPCR, HOXA5 protein expression levels were detected by Western Blot. Clinical data was obtained from inpatient medical records. **Results:** Two microarrays in Oncomine showed that the expression level of HOXA5 was significantly upregulated in AML. Our data revealed that AML patients had higher HOXA5 mRNA and protein expression levels than the controls ($P < 0.001$). The blast percentage in bone marrow of HOXA5 high-expression group was higher than that of HOXA5 low-expression group ($P < 0.05$). Higher expression level of HOXA5 revealed a worse OS in AML

Abbreviations: : SHL, Shuanghuanglian; AML, acute myeloid leukemia; MLL, mixed-lineage leukemia; ESC, embryonic stem cells; HSCT, allogeneic hematopoietic stem cell transplantation; TCGA, The Cancer Genome Atlas; FAB, The French, American, and British; PPI, The protein-protein interaction; TALE, three amino acid loop extension.

* Conflict of interest: The authors declare no conflicts of interests.

* Corresponding to: Wenjun Liu. Department of Pediatrics, Affiliated Hospital of Southwest Medical University, Birth Defects Clinical Medical Research Center of Sichuan, Inpatient building, 16 floors, Luzhou, Sichuan, China.

** Wenzhe Ma. State Key Laboratory of Quality Research in Chinese Medicine, MUST, Building H, Rm. 623a, Avenida Wai Long, Taipa, Macau, China

E-mail addresses: wenjun_liu@swmu.edu.cn (W. Liu), wzma@must.edu.mo (W. Ma).

<https://doi.org/10.1016/j.cuprocancer.2020.100673>

0147-0272/© 2020 Elsevier Inc. All rights reserved.

($P < 0.05$). **Conclusion:** Our findings suggested that HOXA5 might have the potential ability to act as a diagnostic biomarker and potential therapeutic target for AML.

© 2020 Elsevier Inc. All rights reserved.

ARTICLE INFO

Keywords: Acute myeloid leukemia; HOXA5; bioinformatics analysis; prognosis

Introduction

Acute leukemia is the most common hematological malignancy in the adolescent and young adult population. It is an aggressive hematological malignancy and is characterized by abnormal proliferation and differentiation of the immature bone marrow cells.¹ In adults, acute myeloid leukemia (AML) is the most prevalent form of acute leukemia.² Currently, targeted treatment for AML is in routine clinical practice which includes about 10 drugs against specific molecular targets and allogeneic hematopoietic stem cell transplantation (HSCT). However, excluding M3, all other types of AML have problems such as more treatment difficulties, poorer prognosis, higher relapse rate, and lower survival rate. Although the exact causes of AML remain elusive, it is believed that both genetic and environmental factors,³ including previous cancer treatment,⁴ exposure to certain chemicals, smoking and family history⁵ are considered to be the causes of disease progression. Therefore, in addition to ensuring a good living environment, finding the exact genetic-related causes is of great importance for understanding and treating AML.

The *HOX* gene family includes HOXA, B, C, and D. The *HOX* gene encodes a family of transcription factors containing homologous domains and plays a significant role in early embryonic development, including the establishment of cell and tissue characteristics, and the regulation of cell proliferation, differentiation, and survival.⁶ Heterodimers can be formed between MEIS1/PBX and HOX proteins in both a DNA-independent and DNA-dependent manner.⁷ HOXA5 has played important roles in multiple tumors, including gastric cancer,⁸ glioma,⁹ esophageal cancer,¹⁰ and so on. It was reported that *HOXA5* gene was up-regulated in regulate ordinary hematopoietic stem and progenitor cell (HS/PC), and was lowly expressed during terminal differentiation.¹¹ HOXA5 silencing has been reported to reduce cell proliferation and induce apoptosis in patients with AML.¹² However, the relationship between HOXA5 expression in AML and prognosis has been rarely described, so we selected *HOXA5* gene for the further exploration. Here, we provide evidence that HOXA5 is involved with AML based on bioinformatics analysis, mainly by exploring its expression in AML, its correlation with other HOX members, and its relationship with the overall survival of AML.

Materials and methods

Bioinformatics analysis

Oncomine database

The mRNA expression values of HOXA5 in AML and normal organs were analyzed by using the Oncomine platform, which collects publicly available cancer micro-array data. Downloaded the raw data on December 31, 2019, and statistical significance was calculated by Oncomine.

Survival analysis

Downloaded the raw data from Oncomine on December 31, 2019. Based on the mean of HOXA5 expression, patients were divided into high expression of HOXA5 group (HOXA5-high) and low expression of HOXA5 group (HOXA5-low).

STRING database analysis

The STRING database has collected, scored, and integrated all publicly available sources of protein-protein interaction (PPI) information, and supplemented these with computational predictions (<https://string-db.org/>). The goal is to build an exhaustive and objective global network, including physical as well as functional interactions. This study retrieved the data with the following settings: HOXA5; Homo sapiens; high confidence (0.700); the maximum number of interactors displayed: no more than 50 interactions.

Gene coexpression with HOXA5 in TCGA dataset

We used The Cancer Genome Atlas (TCGA) which had both sequencing and pathological data on 30 different cancers. The AML (TCGA, PanCancer Atlas) dataset including data from 173 cases with mRNA expression z-scores (RNA Seq V2 RSEM) was selected for further analyses of HOXA5 using by cBio- Portal (<http://www.cbioportal.org>). Gene coexpression with HOXA5 in PPI network were calculated according to the cBioPortal. The search date was December 31, 2019.

Patients and specimens

Fifty-three newly-diagnosed AML patients (non-M3 subtype) were finally enrolled in Affiliated Hospital of Southwest Medical University from January 2019 to May 2020 were selected, and 19 participants with benign hematological diseases including anemia and thrombocytosis were selected as control group. AML patients were diagnosed based on WHO 2008 criteria and classified according to FAB classification. The treatment of AML patients is mainly based on the Chinese guidelines for diagnosis and treatment of adult AML (not APL) (2017)¹³ at the time of admission. ELN Criteria were used for prognostication and of AML patients. Factors influencing survival were incorporated in the EPI score applicable to adults aged between 15 and 60 years. Poor outcome is associated with shorter CR1 duration, increasing age at the time of relapse, non-favorable karyotype at initial diagnosis, or history of prior allogeneic HCT.¹⁴ A total of 43 mutant genes were detected by next-generation sequencing, including NPM1, N-RAS, c-KIT, TET2, WT1, FLT3, DNMT3A, PTPN11, CBF β -MYH11, AML-ETO, TEL-JAK2, C ABL-BCR, EBPA and p53. According to cytogenetics and molecular genetic standards, patients with non-M3 subtypes of AML are divided into good, moderate and poor prognosis groups. The study was approved by the Affiliated Hospital of Southwest Medical University was conducted in accordance with the Declaration of Helsinki.

Quantitative real-time PCR (RT-qPCR)

Total RNA was extracted using RZ solution (Transgen, China). cDNA was synthesized using a TranScript All-in-One First-Strand cDNA Synthesis SuperMix for qPCR kit (Transgen, China), and TranStart Tip Green Qpcr SuperMix(Transgen, China) was used to assay the HOXA5 mRNA levels. The amplification proceeded as follows: 94°C, 30 seconds, 1 cycle; 94°C, 5 seconds, 60°C, 20 seconds, 40 cycles. The primers were HOXA5- forward, 5'-TTTTGCGGTCGCTATCC-3', HOXA5-reverse, 5'-CTGAGATCCATGCCATTGTAG-3', β -Actin- forward, 5'-GGCGGCACCACCATGTACCC-3', β -Actin - reverse, 5'- CCACACGGAGTACTTGCGC -3'. The $2^{-\Delta\Delta CT}$ method was used to calculate relative mRNA expression. Statistical analysis was performed with SPSS (Version 19.0, IBM Corp. Armonk, NY) and GraphPad Prism (Version 7.01, GraphPad Software). Taking the median as the node, AML patients were divided into HOXA5 high expression group and low expression group. We used Mann-Whitney test to compare differences between 2 groups. It was considered statistically significant when *P* values <0.05.

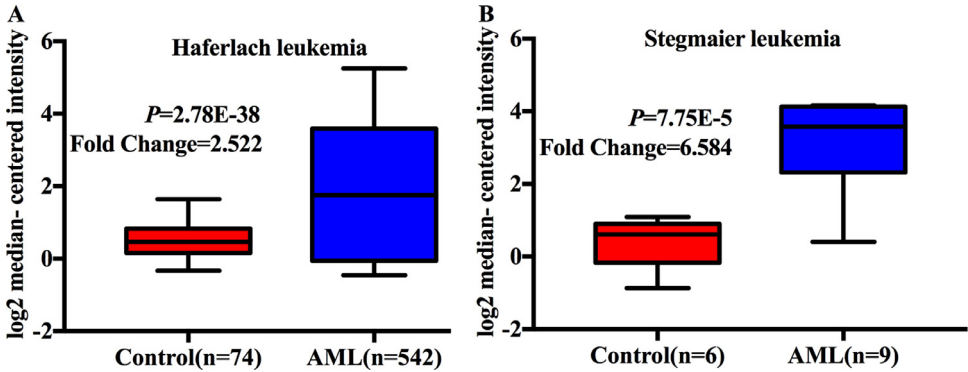


Fig. 1. *HOXA5* gene mRNA expression in AML in the Oncomine leukemia database. (A) *HOXA5* mRNA expression in Haferlach leukemia dataset, analyzed by Oncomine. (B) *HOXA5* mRNA expression in Stegmaier leukemia dataset, analyzed by Oncomine.

Western blotting analyses

BMMCs were lysed in RIPA buffer. Protein concentration was determined using the Bradford dye binding assay with bovine serum albumin as the standard. Total proteins were resolved on SDS–polyacrylamide gel electrophoresis and transferred onto a nitrocellulose membrane. Then, the membrane was incubated with *HOXA5*(1:1000) (abcam, Britain) and antibactin (1:1000) antibodies (CST,USA), followed by incubating with secondary antibodies overnight. The bands were visualized using the enhanced chemiluminescence system (Beijing 4A Biotech Co., Ltd, China).

Statistical analysis

Statistical analysis was performed with GraphPad Prism (Version 7.01, GraphPad Software). We used *t* test or Mann–Whitney test to compare differences between 2 groups. OS was estimated by the Kaplan–Meier method with Log Rank. Univariate analysis, multivariate analysis, hazard ratio and 95% confidence interval were calculated by Cox regression. Based on the mean of *HOXA5* expression, patients were divided into high expression of *HOXA5* group (*HOXA5*-high) and low expression of *HOXA5* group (*HOXA5*-low). It has statistical significance (*P* <0.05).

Results

HOXA5 is highly expressed in acute myeloid leukemia(Oncomine leukemia database and AML patients)

We investigated *HOX* mRNA expression in tumors and it showed that *HOXA5* was higher expressed in leukemia than the corresponding normal tissues (Fig. S1). Next, we analyzed *NAT10* levels in AML patients in our center. In total, 40 of 48 AML patients were non-M3 subtype and 8 were acute promyelocytic leukemia (APL, M3 subtype). *NAT10* was upregulated in AML patients (*P* <0.01, Fig. 1C). There were also no significant differences in *NAT10* expression among different age, gender, or among patients with good, intermediate, or poor prognosis (Table 1). We investigated *HOXA5* expression in AML from Oncomine database including Haferlach leukemia dataset (n = 542, *P* = 2.78E-38, fold change = 2.52)¹⁵ and Stegmaier leukemia dataset (n = 9, *P* = 7.75E-5, fold change = 6.584).¹⁶ The results showed that *HOXA5* levels in AML patients were higher than in the normal ones (*P* <0.01, Fig. 1A and B). Next, we analyzed *HOXA5* levels in

Table 1

Relationships between baseline characteristics and HOXA5 mRNA expression.

			HOXA5-low(%)	HOXA5- high (%)	P value
Age	<60	43	22	21	0.7643 [#]
	≥60	15	7	8	
Gender	Male	34	20 (69%)	20 (69%)	0.1097 [#]
	Female	24	9 (31%)	15 (52%)	
Immunophenotyping	M0	6	2	4	
	M1	4	3	1	
	M2	19	7	12	
	M4	19	13	6	
	M5	10	4	6	
Gene mutation/fusion	FLT3	11	4	7	
	N-RAS	6	2	4	
	c-KIT	2	0	2	
	NPM1	3	1	2	
	WT1	3	1	2	
	IDH1	1	0	1	
	DNMT3A	3	1	2	
	CEBPA	3	1	2	
	AML-ETO	5	2	3	
	TEL-JAK2	3	1	3	
	ABL-BCR	1	0	1	
Risk	Good	5	3 (60%)	2 (40%)	
	Intermediate	14	8 (57%)	8 (57%)	
	Poor	39	18 (46%)	18 (46%)	
Lab examinations	WBC		42.53 ± 10.22	32.79 ± 10.27	0.5042 [*]
	Hb		84.66 ± 5.043	84.17 ± 4.816	0.9451 [*]
	PLT		50.79 ± 11.58	91.03 ± 34.13	0.2689 [*]
	Blast in peripheral blood		53.69 ± 3.368	48.10 ± 4.01	0.3614 [*]
	Blast in bone marrow		41.75 ± 3.572	55.17 ± 4.271	0.0193 [*]
Complete remission		28	16(53%)	12(47%)	

[#]Chi-square test.

AML patients in our hospital. 58 AML patients were non-M3 subtype. NAT10 mRNA and protein were upregulated in AML patients ($P < 0.01$, Fig. 2). The blast percentage in bone marrow had great significant differences in HOXA5 high-expression group and low-expression group (Table 1, Fig. 3). There were also no significant differences in HOXA5 expression among different age, gender, or among patients with good, intermediate, or poor prognosis (Table 1). The white blood cell count, hemoglobin, and thrombocyte count had no differences between HOXA5 high-expression group and low-expression group (Table 1). We also analyzed the relationship between HOXA5 mRNA levels and some mutant genes. However, the number clinical cases of gene mutation and fusion in this study is too small (Table 1), the statistical significance of HOXA5 expression and their relationship is meaningless.

The expression of HOXA5 in AML FAB subtypes

French, American, and British (FAB) classification systems are widely,¹⁷ and traditionally it applied to classify AML as discrete types of M0-M7. The expression of HOXA5 mRNA in different FAB subsets of AML was analyzed in the Oncomine database. Three datasets meet the search criteria were selected for further analysis, including the TCGA dataset (<http://tcga-data.nci.nih.gov/tcga/>), the Valk Leukemia dataset,¹⁸ and the Wouters leukemia dataset.¹⁹ HOXA5 was significantly over-expressed in FAB subtype M5 and under-expressed in FAB subtype M4Eo compared with other FAB subtypes (Fig 4).

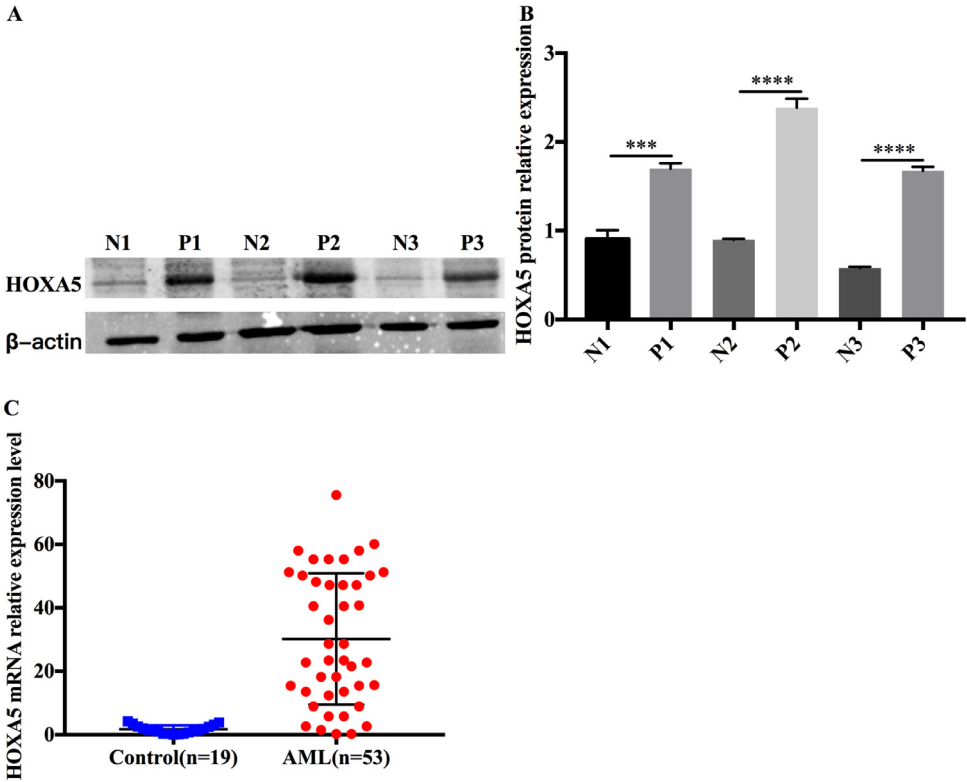


Fig. 2. HOXA5 expression in AML patient (A) HOXA5 protein expression in AML patients and controls was assayed by Western blot. N1, 2, 3: Normal 1, 2, 3; P1,2,3: Patient 1, 2, 3. (B): The histogram of HOXA5 protein in AML and controls. Statistical significance was calculated by. (** <0.0005 , **** <0.0001 , t-Test). (C) HOXA5 mRNA expression in AML patients and controls was assayed by RT-qPCR (** $^*P < 0.0001$, t test).

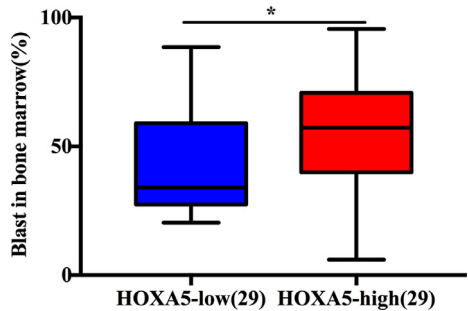


Fig. 3. The blast percentage in bone marrow of HOXA5 high-expression group and low-expression group. Statistical significance was calculated by t test. (* <0.05).

High level of HOXA5 is associated with poor prognoses in AML patients

HOXA5 is highly expressed in AML. To further analysis whether it could be a prognostic marker for AML, 4 datasets available with the clinical outcome information were selected for survival analysis with the Kaplan-Meier plotter. As shown in Fig. 5A, the increased HOXA5 mRNA is significantly correlated with shorter overall survival time of AML patients in the TCGA dataset

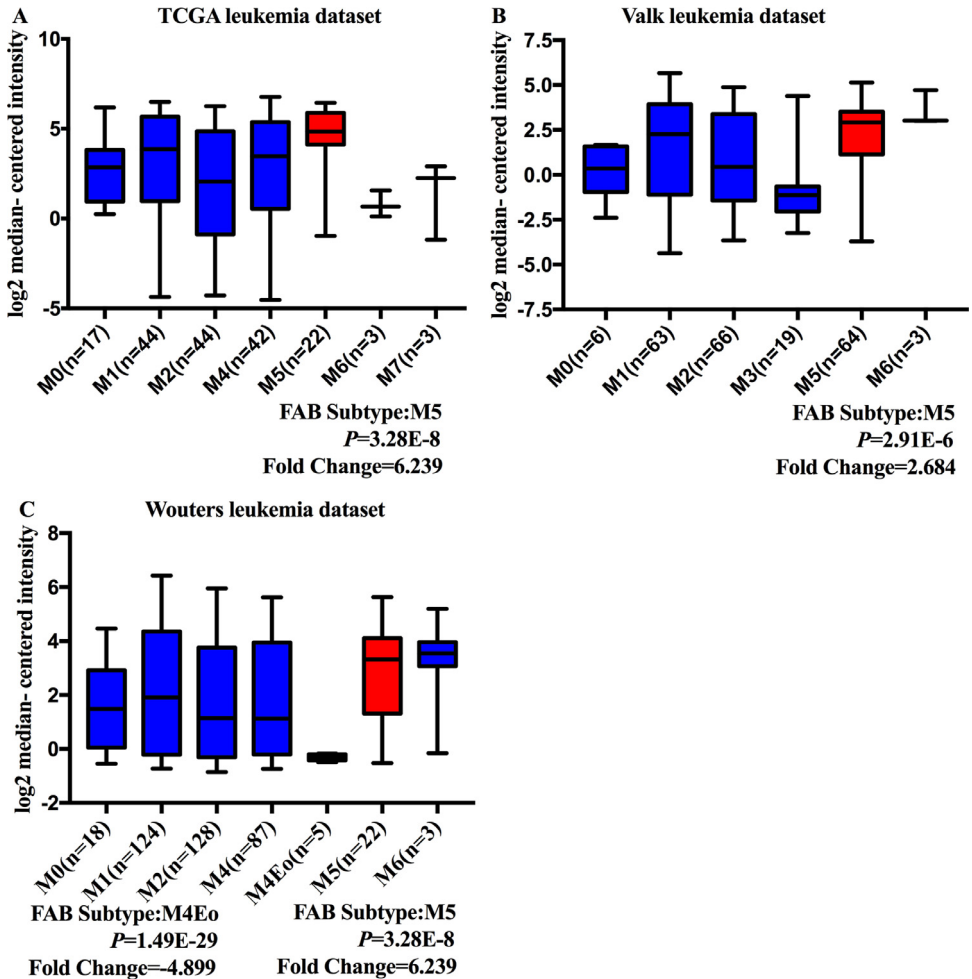


Fig. 4. HOXA5 mRNA expression in AML FAB subtypes. (A) HOXA5 mRNA expression in TCGA leukemia dataset, analyzed by OncoPrint. (B) HOXA5 mRNA expression in Valk leukemia dataset, analyzed by OncoPrint. (C) HOXA5 mRNA expression in Wouters leukemia dataset, analyzed by OncoPrint.

($P=0.0405$) (<http://tcga-data.nci.nih.gov/tcga/>). The similar trends, although not significant, were observed in the Metzeler leukemia dataset²⁰, and the Rapini leukemia dataset (Fig. 5B–C).²¹ However, it was opposite in the Heuser leukemia dataset (Fig. 5D),²² which may be due to the small sample size ($n=28$). Taken together, our data imply that HOXA5 is a potential prognostic marker for AML patients with poor outcome.

The PPI network of HOXA5 protein

To achieve a comprehensive and objective global PPI network, HOXA5 was analyzed in the STRING database. The PPI enrichment P -value was $< 1.0E-16$ and the number of nodes was 23. Not unexpectedly, HOXA5 interacted with most other HOX family genes, including HOXA3/6/7/9, HOXB3/4/5/6/7/8, HOXC4/5/6/8/9 and HOXD4/9 through the co-expression, gene fusion, and gene co-occurrence manner (Fig. 6). HOXA5 also interacted with transcriptional

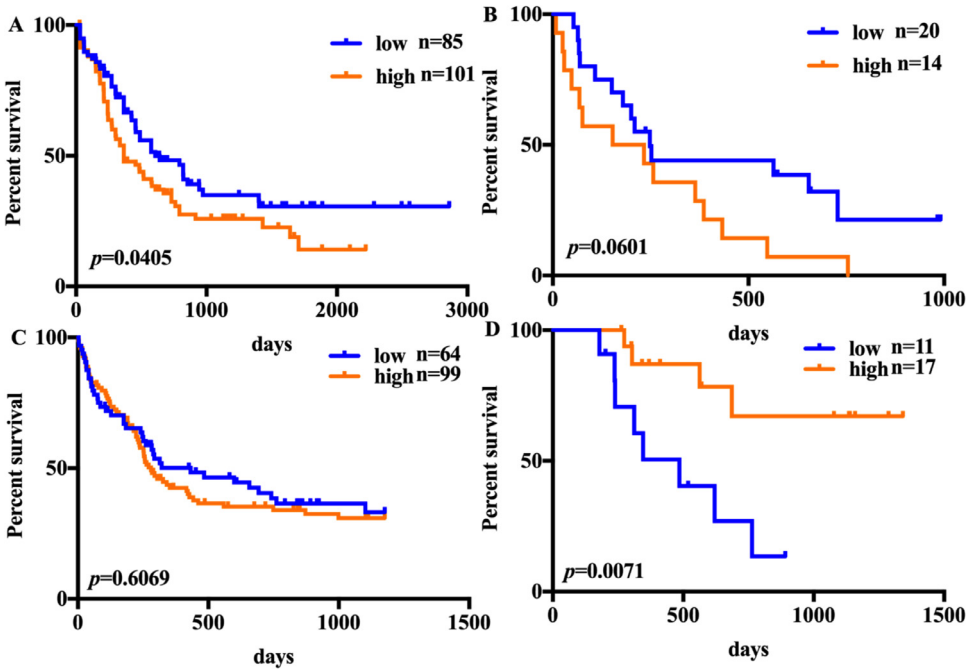


Fig. 5. The prognostic value of HOXA5 mRNA level in AML patients (OS in Kaplan-Meier plotter). (A)TCGA dataset; (B)Raponi leukemia dataset; (C) Metzeler leukemia dataset; (D)Heuser leukemia dataset. It has statistical significance ($P < 0.05$).

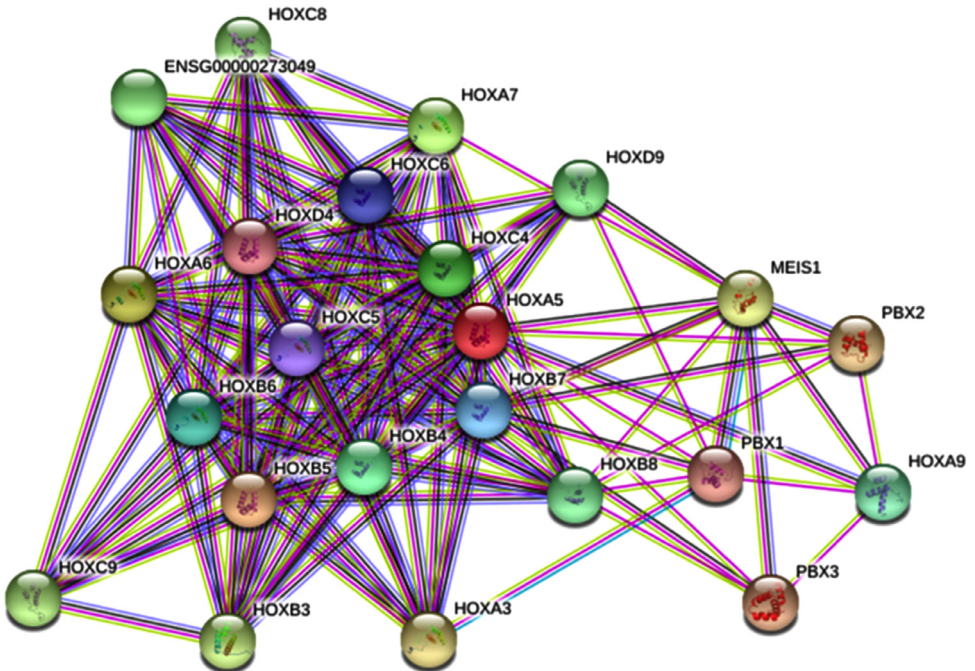


Fig. 6. HOXA5 PPI network complex and modular analysis.

cofactors PBX1/2/3 and MEIS1, which belong to the highly conserved TALE (“three amino acid loop extension”) family of non-HOX HD cofactors and are required for the HOX family proteins to form a synergistic DNA-binding complex.²³ PBX cofactors supervise and guarantee normal hematopoiesis, the pluripotency of embryonic stem cells (ESC) and tissue regeneration, similar to HOX proteins in hematopoiesis.²⁴ Meis1 is highly expressed in mixed-lineage leukemia (MLL) and plays an important role in establishing leukemic stem cell potential and frequency by quantitatively regulating the extent of self-renewal, differentiation arrest, and cycling.²⁵ The interactions of HOXA5 with other HOX proteins and the TALE cotranscriptional factors were further proved by the coexpression in AML patients (Fig. S3)), implying the potential role of HOXA5 in AML pathogenesis.

Known associations: blue lines indicate protein interactions confirmed by the source database, pink lines indicate experimentally validated protein interaction relationships. Predicted associations: green lines refer to gene neighbors, red lines refer to gene fusion, purple lines refer to gene co-occurrence. others: cyan lines refer to protein associations by text mining, black lines represent co-expression, and lavender represents protein homology.

Discussion

HOX gene disorder play a significant role in leukemogenesis, and among the family members, HOXA5 is less researched. Here, by bioinformatics analysis, we highlighted the importance of HOXA5 in AML. We found that HOXA5 mRNA was differentially expressed in normal and tumor tissues (Fig. S1). However, it was significantly overexpressed in AML patients compared to normal subjects' peripheral blood mononuclear cell, monocytes or neutrophils. The increased HOXA5 mRNA was significantly associated with shorter over survival time in AML patients. It is true that some of the data reported in our study are actually conflicting (Fig 5). Out of 4 datasets used, 2 gave no differences but the similar trends were observed. The other 2 datasets indeed showed opposite results. But the number of sample in the TCGA dataset (n = 186) and the Heuser leukemia dataset (n = 28) had great difference. The opposite result in the Heuser leukemia dataset may be due to the small sample size.

PPI network analysis indicated that HOXA5 was closely co-expressed, fused or interacted with the other HOX family members and transcriptional cofactors, most of which have been reported in the development AML.²⁶

The main feature of AML is the abnormal proliferation of primitive and naive bone marrow cells in peripheral blood. The main clinical characteristics are anemia, bleeding, infection and fever, organ infiltration, abnormal metabolism, and so on. AML contributes to 15%-20% of the childhood leukemias.²⁷ Although the outcome has improved significantly in the past few decades, AML is still a malignant tumor that threatens the lives of children, with current OS rate of ~70%.²⁸ While, complete remission (CR) is achieved in 75%-80% of adult patients.¹⁴ Whether children or adults, relapse is still the most common cause of treatment failure and the main obstacle to further improve the prognosis of AML.^{29,30} Our finding is consistent with the previous report that patients with AML who had high expression of HOXA5 had significantly lower complete remission rates.³¹ The 3-year overall survival rate in AML patients with high expression level of HOXA5 was much lower than that of the low populations (40.5% vs 82.5%).³² It was also reported that the suppression of the expression of HOXA5 by short hairpin RNA can suppressed the proliferation of AML cells and enhanced the sensitivity of AML cells to cytarabine.¹² We found that HOXA5 was significantly over-expressed in FAB subtype M5 and under-expressed in FAB subtype M4Eo compared with other FAB subtypes (Fig 4). The subtype of M4eo also has better prognosis than other subgroups.³³ So, our study and the published data suggested that HOXA5 may be a potential prognostic biomarker of AML patients.

How HOXA5 is regulated in AML cells? Gene fusion and mutation are a common event in the development of leukemia, especially in the development of AML.³⁴ Nucleoporin 98 gene (NUP98) is able to fuse with NSD1, JARID1A, PHF23, and HOXA9/10/13, which has been reported to be associated with increased HOXA5 expression.³⁵ It has been reported that CALM-AF10 can enhance

the expression of HOXA5 by the regulation of H3K79 methylation,³⁶ which plays important roles in leukemogenesis.³⁷ Promoter methylation is another notable mechanism that regulates HOX mRNA expression and is generally negatively related to the expression of the gene.³⁸ However, it was reported that the HOXA5 promoter is highly methylated in more than 60% of AML samples.³⁹ This indicated that more than 60% of AML samples had low expression of HOXA5, which is contrary to our finding. It is probably due to the small sample size ($n=25$) in this study. Aberrant HOX expression is found in nearly all AMLs that harbor a mutation in the Nucleophosmin (NPM1) gene, and FLT3 is concomitantly mutated in approximately 60% of these cases.⁴⁰ It was reported that the expression of HOXA5 mRNA level in AML patients with NPM1 mutations was high.^{26,41} This may be caused by that NPM1 mutations result in abnormal cytoplasmic localization of the mutant protein (NPM1c) in AML cells; NPM1 mutation can immediately downregulate the expression of HOX family genes followed by differentiation.⁴² However, in the previous study, the expression of HOXA5 in AML cells with NPM1 mutation was very low.⁴³ The reports are not entirely consistent. In addition, Previous study has reported that leukemia can be alone induced by overexpression of individual HOXA genes.⁴⁴ Study has found that NPMc+ leukemic cell survival required upregulation of HOX genes.⁴⁵ As reported, NPM1 mutation is an indicator of good prognosis for AML.⁴⁶ However, the study found the simultaneous presence of NPM1 mutation and high HOXA5 expression in AML patients had worse overall survival.²⁶ Mutations of the FMS-like tyrosine kinase 3 (FLT3) gene is a common driver mutation that presents with a high leukemic burden and confers a poor prognosis in patients with AML.⁴⁷ Flt3-ITD(+) AML bone marrow samples highly expressed HOXA5 gene.⁴⁸ In our study, we found that the number of FLT3 mutations was higher in HOXA5-high expression than HOXA5-low expression. However, HOXA gene can regulate Flt3 in hematopoietic Progenitors.⁴⁹ FLT3 mutation had no significant effect on the expression of HOXA5.⁵⁰ It may be caused by the small number of the AML samples. There were not many cases of other mutations and gene fusion in the samples we collected (Table 1). But the complicated relationship among HOXA5, gene fusion and mutation in AML still needs to be explored in depth.

Taken together, this evidence implies that HOXA5 plays important roles in the pathogenesis of AML. However, because there are few clinical cases of gene mutation and fusion in this study, the statistical significance of HOXA5 expression and their relationship is meaningless. The detailed mechanisms and whether it could be a therapeutic target for AML still need further investigations.

Conclusions

In summary, the increased HOXA5 was associated with significantly shorter overall survival time in AML patients. This study indicates that HOXA5 might be a prognostic marker and a potential therapeutic target for AML.

Ethics committee approval

The study was approved by the Affiliated Hospital of Southwest Medical University was conducted in accordance with the Declaration of Helsinki (No. KY2020110).

Acknowledgments

This work was funded by the Science and Technology Development Fund, Macau SAR (File no. 0013/2019/A1), Basic Research Project of Sichuan Province, China (File No. 2019YJ0690), The Major Science and Technology Projects in Sichuan Province (No. 2019YFS0531), Research Subject of

Sichuan Provincial Health and Health Committee, China (File No. [18PJ035](#)), The Affiliated Hospital of Southwest Medical University, Luzhou, Sichuan, China (File No. [2017-PT-30](#)), The Southwest Medical University, Luzhou, Sichuan, China (File No. [2017-ZRQN-149](#)).

Supplementary materials

Supplementary material associated with this article can be found, in the online version, at doi:[10.1016/j.cupr.2020.100673](#).

References

- Juliussøn G, Hough R. Leukemia. *Prog Tumor Res.* 2016;43:87–100.
- Shallis RM, Wang R, Davidoff A, Ma XM, Zeidan AM. Epidemiology of acute myeloid leukemia: recent progress and enduring challenges. *Blood Rev.* 2019;36:70–87.
- Abelson S, Collord G, Ng SWK, et al. Prediction of acute myeloid leukaemia risk in healthy individuals. *Nature.* 2018;559:400–404.
- Shenolikar R, Durden E, Meyer N, Lenhart G, Moore K. Incidence of secondary myelodysplastic syndrome (MDS) and acute myeloid leukemia (AML) in patients with ovarian or breast cancer in a real-world setting in the United States. *Gynecol Oncol.* 2018;151:190–195.
- Sandner AS, Weggel R, Mehraein Y, Schneider S, Hiddemann W, Spiekermann K. Frequency of hematologic and solid malignancies in the family history of 50 patients with acute myeloid leukemia - a single center analysis. *PLoS One.* 2019;14.
- Primon M, Hunter KD, Pandha HS, et al. Kinase regulation of HOX transcription factors. *Cancers (Basel).* 2019;11:508.
- Bisaillon R, Wilhelm BT, Kros J, Sauvageau G. C-terminal domain of MEIS1 converts PKNX1 (PREP1) into a HOXA9-collaborating oncoprotein. *Blood.* 2011;118:4682–4689.
- Wu YX, Zhou T, Tang Q, Xiao JW. HOXA5 inhibits tumor growth of gastric cancer under the regulation of microRNA-196a. *Gene.* 2019;681:62–68.
- Cimino PJ, Kim Y, Wu HJ, et al. Increased HOXA5 expression provides a selective advantage for gain of whole chromosome 7 in IDH wild-type glioblastoma. *Genes Dev.* 2018;32:512–523.
- Zhang H, Zhao JH, Suo ZM. Knockdown of HOXA5 inhibits the tumorigenesis in esophageal squamous cell cancer. *Biomed Pharmacother.* 2017;86:149–154.
- Ketty LM, Lebert-Ghali CE, Grishagin IV, et al. Pathways, processes, and candidate drugs associated with a *Hoxa* cluster-dependency model of leukemia. *Cancers (Basel).* 2019;11:2036.
- Li N, Jia XH, Wang JY, Li YJ, Xie SY. Knockdown of homeobox A5 by small hairpin RNA inhibits proliferation and enhances cytarabine chemosensitivity of acute myeloid leukemia cells. *Mol Med Rep.* 2015;12:6861–6866.
- Wei H. Chinese guidelines for diagnosis and treatment of adult myeloid leukemia (Not APL) (2017). *Chin J Hematol.* 2017;38:177–182.
- Dohner H, Estey E, Grimwade D, et al. Diagnosis and management of AML in adults: 2017 ELN recommendations from an international expert panel. *Blood.* 2017;129:424–447.
- Haeflrich T, Kohlmann A, Wiczorek L, et al. Clinical utility of microarray-based gene expression profiling in the diagnosis and subclassification of leukemia: report from the International Microarray Innovations in Leukemia Study Group. *J Clin Oncol.* 2010;28:2529–2537.
- Stegmaier K, Ross KN, Colavito SA, O'Malley S, Stockwell BR, Golub TR. Gene expression-based high-throughput screening (GE-HTS) and application to leukemia differentiation. *Nat Genet.* 2004;36:257–263.
- Canaani J, Beohou E, Labopin M, et al. Impact of FAB classification on predicting outcome in acute myeloid leukemia, not otherwise specified, patients undergoing allogeneic stem cell transplantation in CR1: an analysis of 1690 patients from the acute leukemia working party of EBMT. *Am J Hematol.* 2017;92:344–350.
- Valk PJM, Verhaak RGW, Beijnen MA, et al. Prognostically useful gene-expression profiles in acute myeloid leukemia. *N Engl J Med.* 2004;350:1617–1628.
- Wouters BJ, Lowenberg B, Erpelinck-Verschueren CA, van Putten WL, Valk PJ, Delwel R. Double CEBPA mutations, but not single CEBPA mutations, define a subgroup of acute myeloid leukemia with a distinctive gene expression profile that is uniquely associated with a favorable outcome. *Blood.* 2009;113:3088–3091.
- Metzeler KH, Hummel M, Bloomfield CD, et al. An 86-probe-set gene-expression signature predicts survival in cytogenetically normal acute myeloid leukemia. *Blood.* 2008;112:4193–4201.
- Raponi M, Lancet JE, Fan HT, et al. A 2-gene classifier for predicting response to the farnesyltransferase inhibitor tipifarnib in acute myeloid leukemia. *Blood.* 2008;111:2589–2596.
- Heusser M, Wingen LU, Steinemann D, et al. Gene-expression profiles and their association with drug resistance in adult acute myeloid leukemia. *Haematologica - Hematol J.* 2005;90:1484–1492.
- Blasi F, Bruckmann C, Penkov D, Dardaei L. A tale of TALE, PREP1, PBX1, and MEIS1: interconnections and competition in cancer. *Bioessays.* 2017;39.
- Merabet S, Mann RS. To be specific or not: the critical relationship between Hox and TALE proteins. *Trends Genet.* 2016;32:334–347.
- Yuan XJ, Braun T. An unexpected switch: regulation of cardiomyocyte proliferation by the Homeobox gene Meis1. *Circ Res.* 2013;113:245–248.
- Nagy A, Osz A, Budczies J, et al. Elevated HOX gene expression in acute myeloid leukemia is associated with NPM1 mutations and poor survival. *J Adv Res.* 2019;20:105–116.

27. Reedijk AMJ, Klein K, Coebergh JWW, et al. Improved survival for children and young adolescents with acute myeloid leukemia: a Dutch study on incidence, survival and mortality. *Leukemia*. 2019;33:1349–1359.
28. Creutzig U, van den Heuvel-Eibrink MM, Gibson B, et al. Diagnosis and management of acute myeloid leukemia in children and adolescents: recommendations from an international expert panel. *Blood*. 2012;120:3187–3205.
29. Karlsson L, Forestier E, Hasle H, et al. Outcome after intensive reinduction therapy and allogeneic stem cell transplant in paediatric relapsed acute myeloid leukaemia. *Br J Haematol*. 2017;178:592–602.
30. Yilmaz M, Wang F, Loghavi S, et al. Late relapse in acute myeloid leukemia (AML): clonal evolution or therapy-related leukemia? *Blood Cancer J*. 2019;9:7.
31. Zhao P, Tan L, Ruan J, et al. Aberrant expression of HOXA5 and HOXA9 in AML. *Asian Pac J Cancer Prev*. 2015;16:3941–3944.
32. Kim SY, Hwang SH, Song EJ, Shin HJ, Jung JS, Lee EY. Level of HOXA5 hypermethylation in acute myeloid leukemia is associated with short-term outcome. *Korean J Lab Med*. 2010;30:469–473.
33. Klein K, de Haas V, Bank IEM, et al. Clinical and prognostic significance of eosinophilia and inv(16)/t(16;16) in pediatric acute myelomonocytic leukemia (AML-M4). *Pediatr Blood Cancer*. 2017;64:e26512.
34. Desai P, Mencia-Trinchant N, Savenkov O, et al. Somatic mutations precede myeloid leukemia years before diagnosis. *Nat Med*. 2018;24:1015.
35. Gough SM, Slape CI, Aplan PD. NUP98 gene fusions and hematopoietic malignancies: common themes and new biologic insights. *Blood*. 2011;118:6247–6257.
36. Okada Y, Jiang Q, Lemieux M, Jeannotte L, Su LS, Zhang Y. Leukaemic transformation by CALM-AF10 involves upregulation of Hoxa5 by hDOT1L. *Nat Cell Biol*. 2006;8:1017–U105.
37. Conway AE, Haldeman JM, Wechsler DS, Lavau CP. A critical role for CRM1 in regulating HOXA gene transcription in CALM-AF10 leukemias. *Leukemia*. 2015;29:423–432.
38. da Silva RA, Fuhler GM, Janmaat VT, et al. HOXA cluster gene expression during osteoblast differentiation involves epigenetic control. *Bone*. 2019;125:74–86.
39. Strathdee G, Sim A, Soutar R, Holyoake TL, Brown R. HOXA5 is targeted by cell-type-specific CpG island methylation in normal cells and during the development of acute myeloid leukaemia. *Carcinogenesis*. 2007;28:299–309.
40. Kuhn MWM, Song E, Feng ZH, et al. Targeting chromatin regulators inhibits leukemogenic gene expression in NPM1 mutant leukemia. *Cancer Discovery*. 2016;6:1166–1181.
41. Musialik E, Bujko M, Kober P, et al. Promoter DNA methylation and expression levels of HOXA4, HOXA5 and MEIS1 in acute myeloid leukemia. *Mol Med Rep*. 2015;11:3948–3954.
42. Brunetti L, Gundry MC, Sorcini D, et al. Mutant NPM1 maintains the leukemic state through HOX expression. *Cancer Cell*. 2018;34:499.
43. Ghasemi R, Struthers H, Wilson ER, Spencer DH. Contribution of CTCF binding to transcriptional activity at the HOXA locus in NPM1-mutant AML cells. *Leukemia*.
44. Bach C, Buhl S, Mueller D, Garcia-Cuellar MP, Maethner E, Slany RK. Leukemogenic transformation by HOXA cluster genes. *Blood*. 2010;115:2910–2918.
45. Zhang W, Zhao C, Zhao JM, et al. Inactivation of PBX3 and HOXA9 by down-regulating H3K79 methylation represses NPM1-mutated leukemic cell survival. *Theranostics*. 2018;8:4359–4371.
46. Heath EM, Chan SM, Minden MD, Murphy T, Shlush LI, Schimmer AD. Biological and clinical consequences of NPM1 mutations in AML. *Leukemia*. 2017;31:798–807.
47. Daver N, Schlenk RF, Russell NH, Levis MJ. Targeting FLT3 mutations in AML: review of current knowledge and evidence. *Leukemia*. 2019;33:299–312.
48. Moore MAS, Dorn DC, Schuringa JJ, Chung KY, Morrone G. Constitutive activation of Flt3 and STAT5A enhances self-renewal and alters differentiation of hematopoietic stem cells. *Exp Hematol*. 2007;35:105–116.
49. Gwin K, Frank E, Bossou A, Medina KL. Hoxa9 regulates Flt3 in lymphohematopoietic progenitors. *J Immunol*. 2010;185:6572–6583.
50. Skvarova Kramarzova K, Fiser K, Mejstrikova E, et al. Homeobox gene expression in acute myeloid leukemia is linked to typical underlying molecular aberrations. *J Hematol Oncol*. 2014;7:94.